Low Resolution Refinement

Phenix Workshop

University of Vienna, May 2018

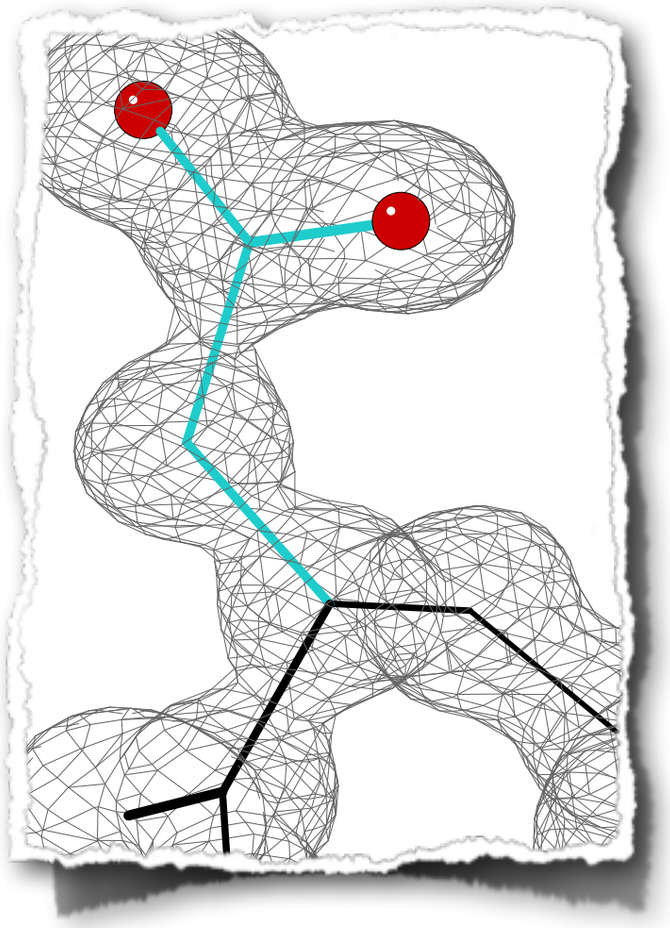
Paul Adams

Lawrence Berkeley Laboratory and

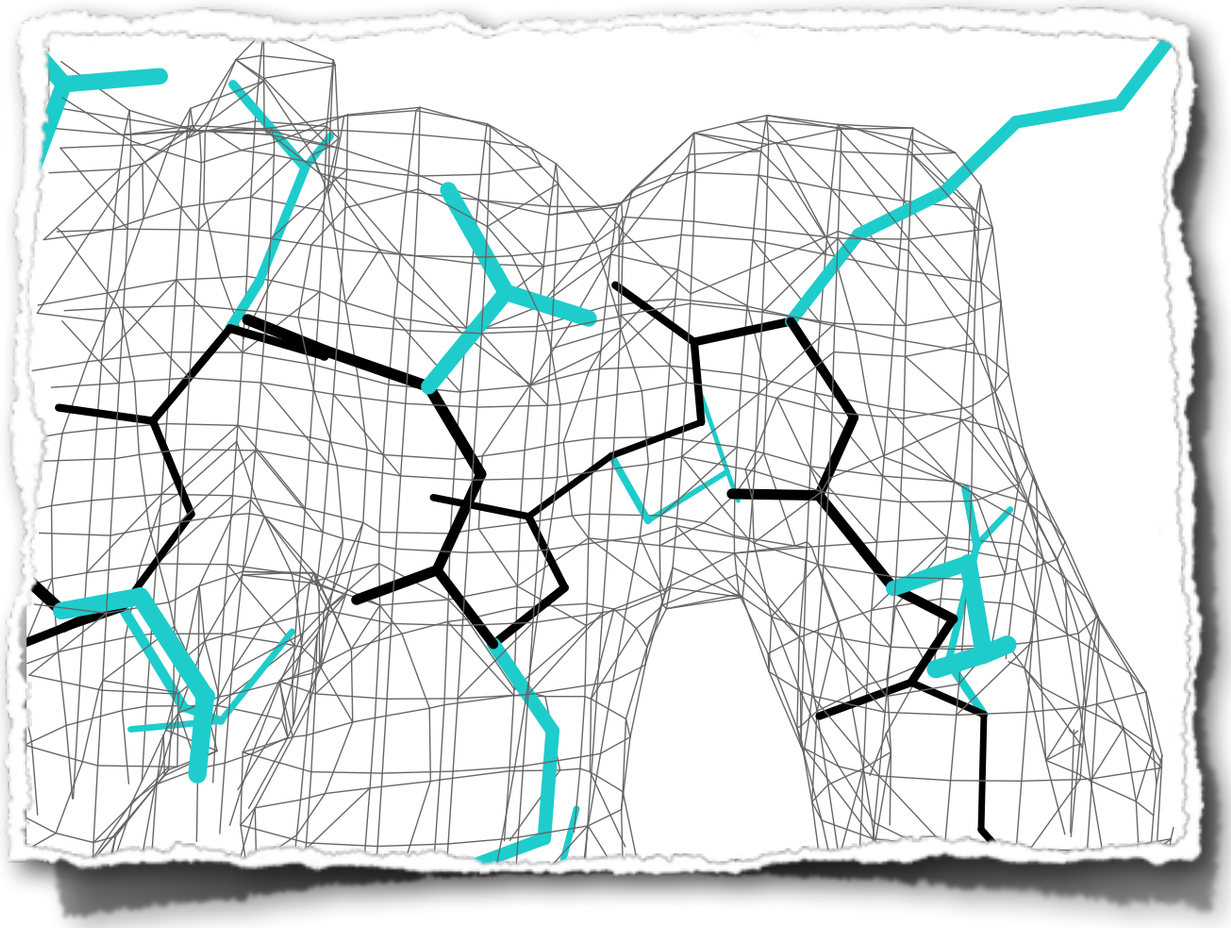
Department of Bioengineering UC Berkeley

Macromolecular Crystallography

PDBID: 2gkg
Resolution: 1.00Å



PDB ID: 3k7a
Resolution: 3.80Å



- Many challenges, but low resolution data is increasingly an issue:
  - How to interpret "featureless" maps (pattern matching, chemical constraints)
  - How to optimize models with sparse data (prior information)

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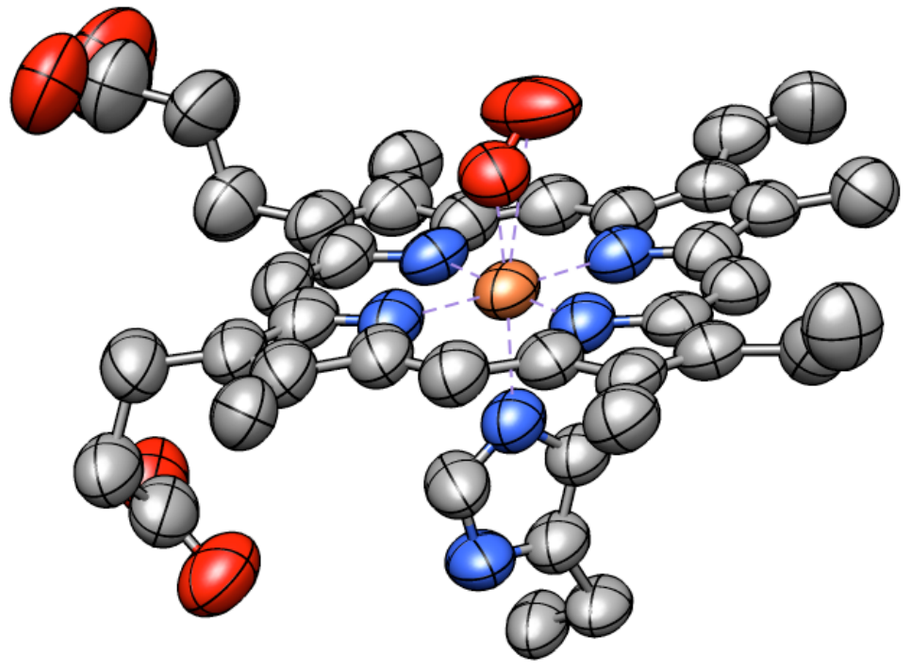
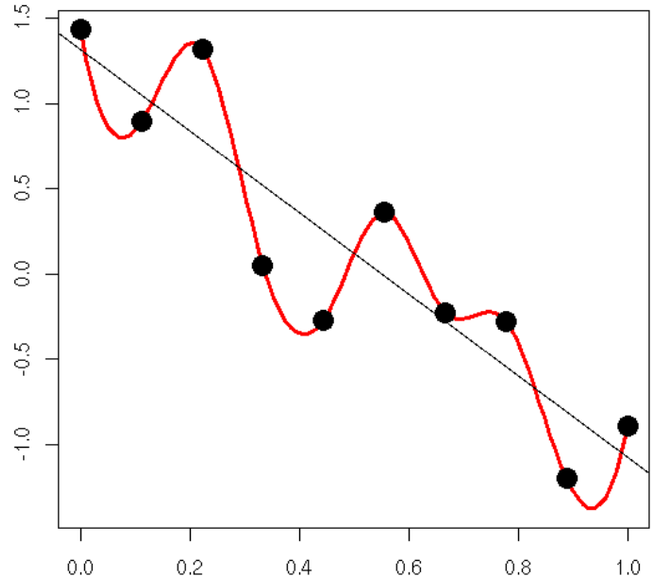
The Challenge of Too Few Data

- With only low resolution data we typically have too many parameters to optimize
  - Atomic coordinates, displacement parameters
- Underdetermined optimization problems lead to overfitting of the data
- To help address overfitting we can:
  - Add prior information to reduce the number of effective parameters
  - Remove parameters
- Current refinement methods do not define a reasonable chemical result in the absence of data





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Improving the Observation to Parameter Ratio

- To make refinement practical the observation to parameter ratio is increased using restraints and constraints:
- Restraint
  - Model property ~ ideal value
  - Adds prior observed information (reduces the number of parameters refined)
  - Inclusion of chemical information in the objective function
- Constraint
  - Model property = ideal value
  - Removes one or more parameters from the model

Methods in Phenix for Improving Models

- Using prior structural knowledge as additional restraints:
  - Secondary structure
  - Protein mainchain conformations (Ramachandran)
  - Related high resolution structures as restraints
  - Multiple copies of the same molecule as restraints *(c.f. local NCS restraints in SHELX)*
- Automated correction of models during refinement using prior knowledge of stereochemistry:
  - Fixing of rotamers
  - Flipping of side chains

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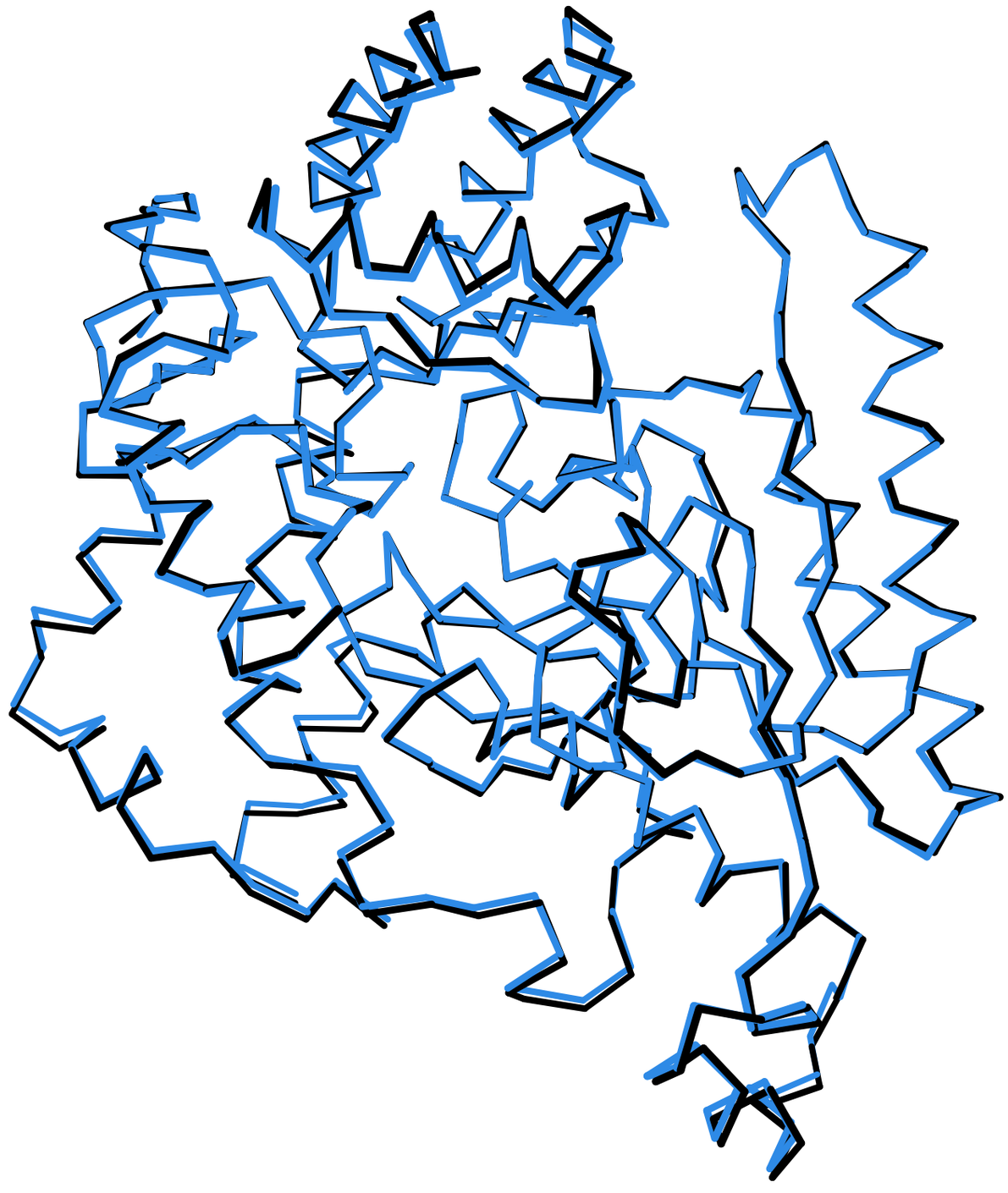
Reference model restraints

(Jeff Headd)

1GTX and 1OHV

1GTX: 3.0 Å

1OHV: 2.3 Å



4-aminobutyrate-aminotransferase

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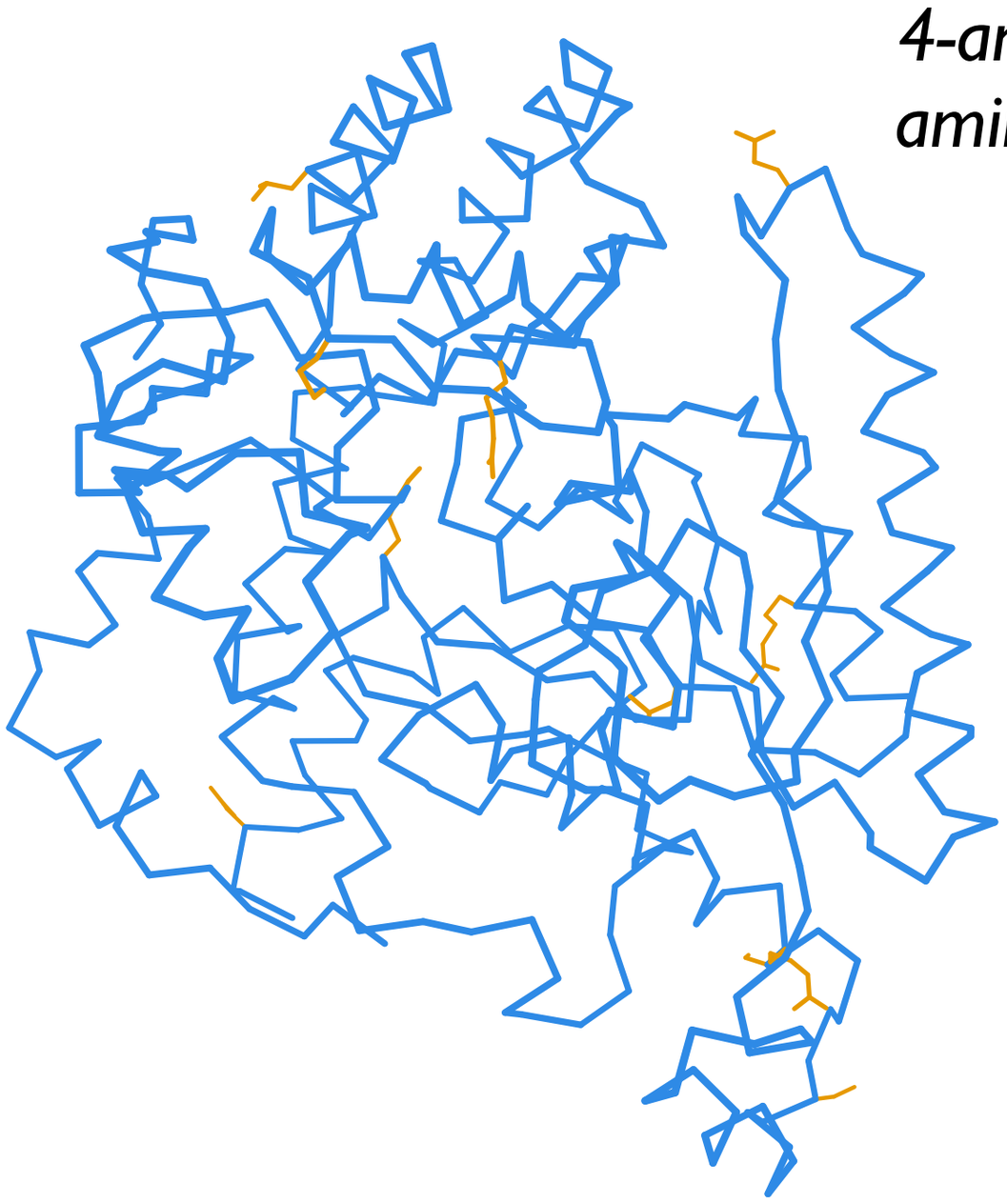
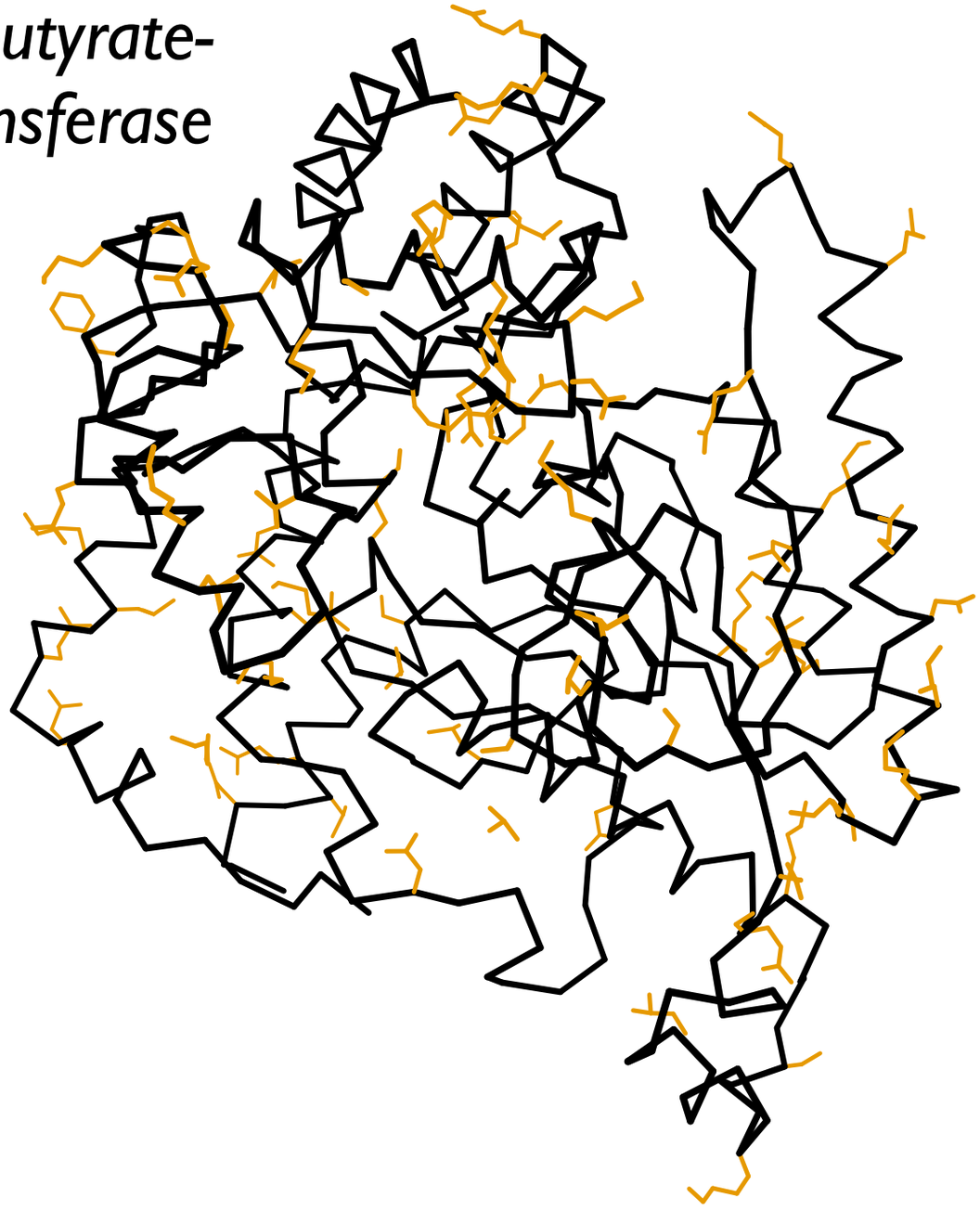
1GTX and 1OHV

4-aminobutyrate-aminotransferase





1OHV: 2.3 Å

1GTX: 3.0 Å

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Reference Model Restraints

Combines two concepts:

- Pre-correct rotamer outliers
  - Set rotamer outliers in the model to match the torsion angles of the reference model if the reference model has an acceptable rotamer at that position and there is no significant clash or density mismatch
- Generate reference torsion restraints
  - Restrain each torsion angle in the working model to the corresponding torsion angle in the reference model
    - Chains are aligned using SSM alignment to allow for sequence differences
  - Restraints take the form of a modified harmonic 'top-out' potential that allows for structural differences

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Headd JJ et al., 2012, *Acta Cryst.* **D68**:381-390

Reference model restraints

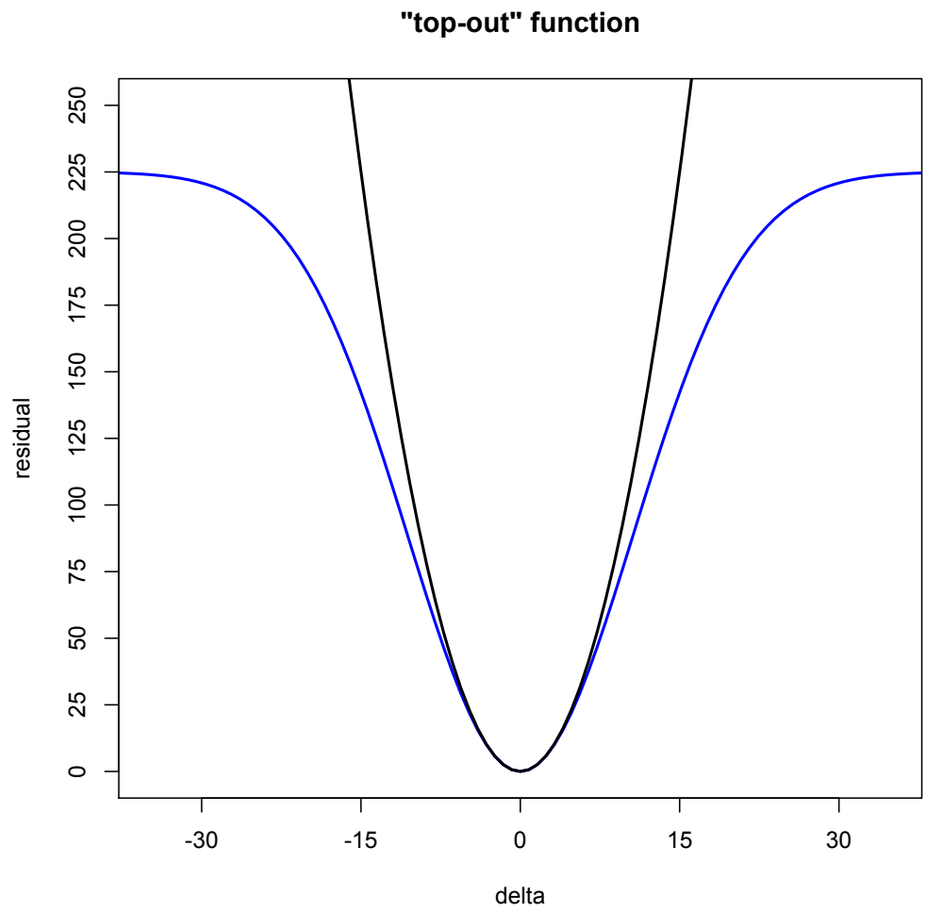
$$E_{total} = \sum_{i=1}^{n} E_i$$

Simple harmonic potential: $E_i = w\Delta_i^2$

'Top-out' potential: $E_i = \tau(1.0 - e^{\frac{-\Delta_i^2}{l^2}})$

$$\tau = wl^2$$

$$w = \frac{1}{\sigma^2}$$



Similar potentials are used in REFMAC5 and BUSTER - Geman-McClure robust estimator function

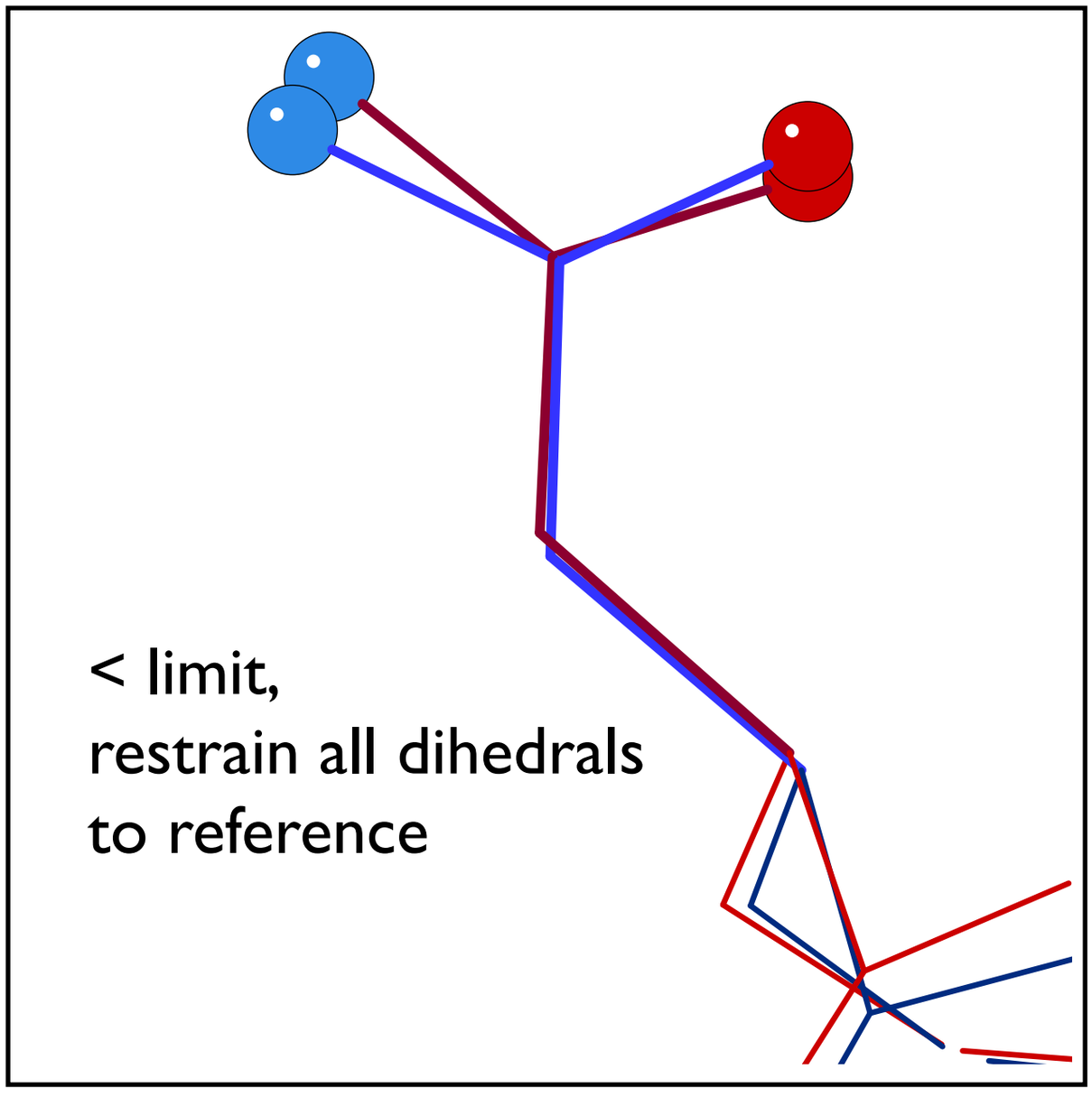
where σ is the ESD, Δ is the difference between the model dihedral and reference dihedral, and *l* is a 'limit' parameter that limits how far the model dihedral may vary from the reference dihedral before being shut off.

developed by Ralf Grosse-Kunstleve

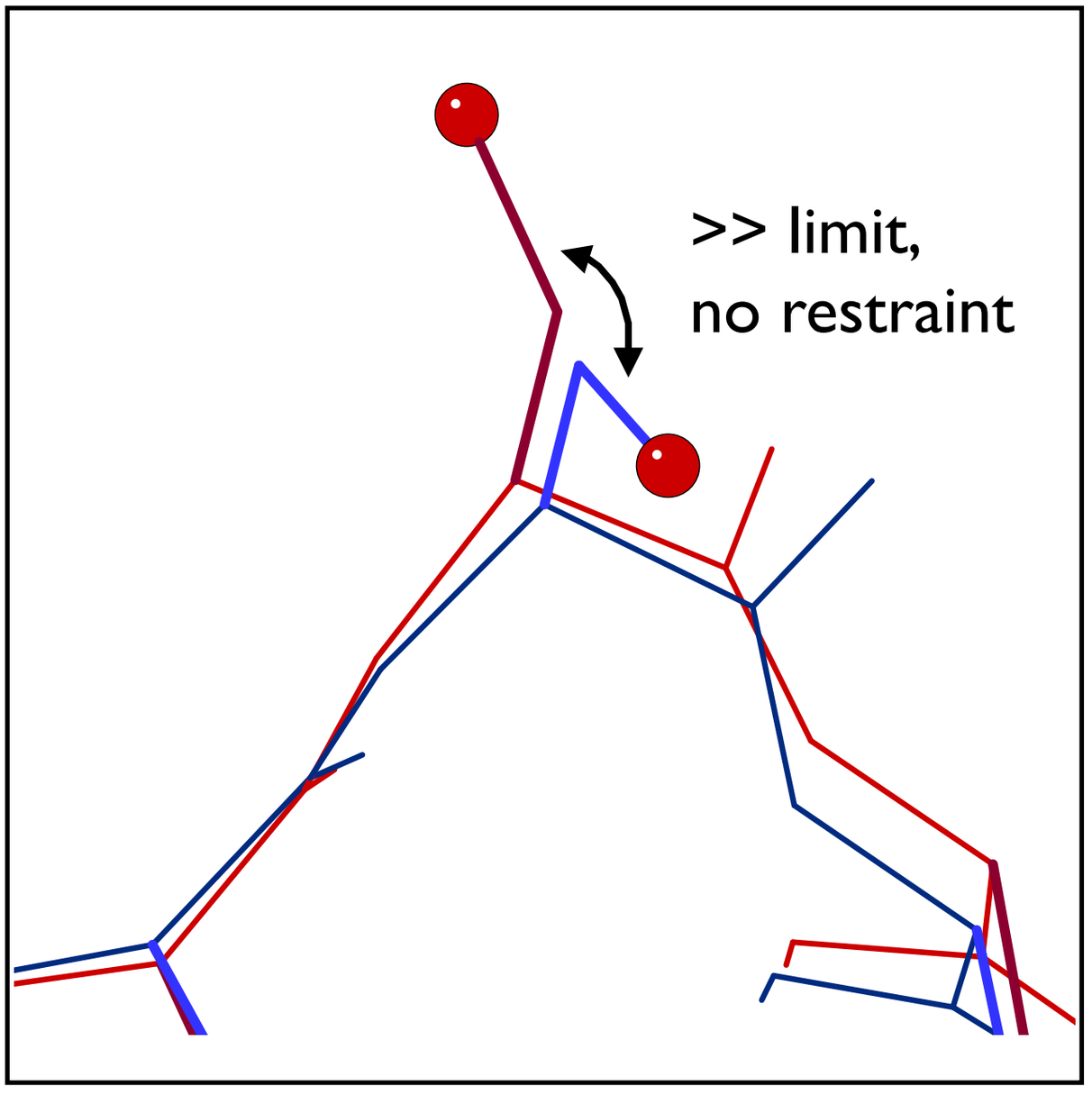
default: limit = 15.0°

The ‘limit’ parameter

default: limit = 15.0°



< limit,
restrain all dihedrals
to reference



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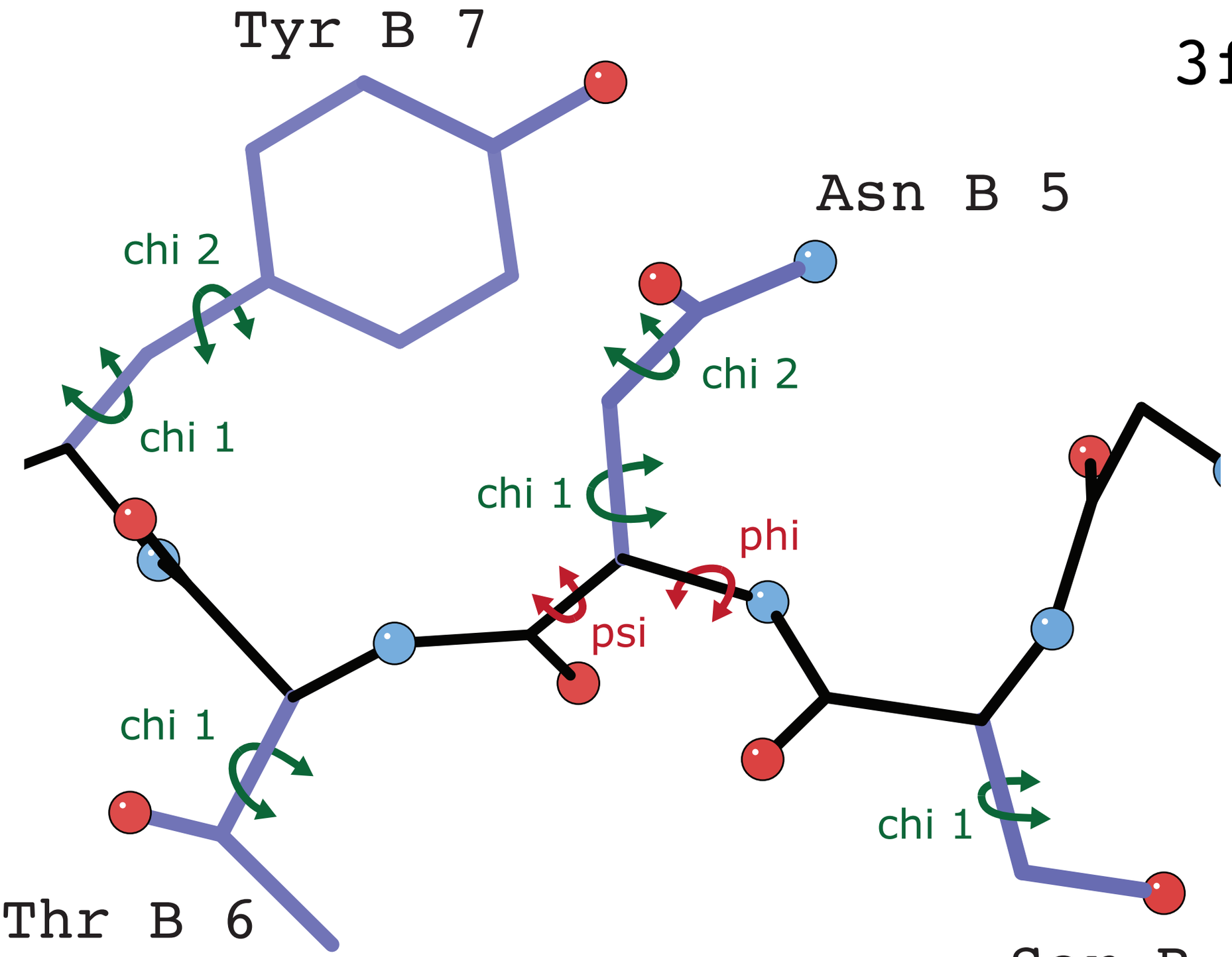
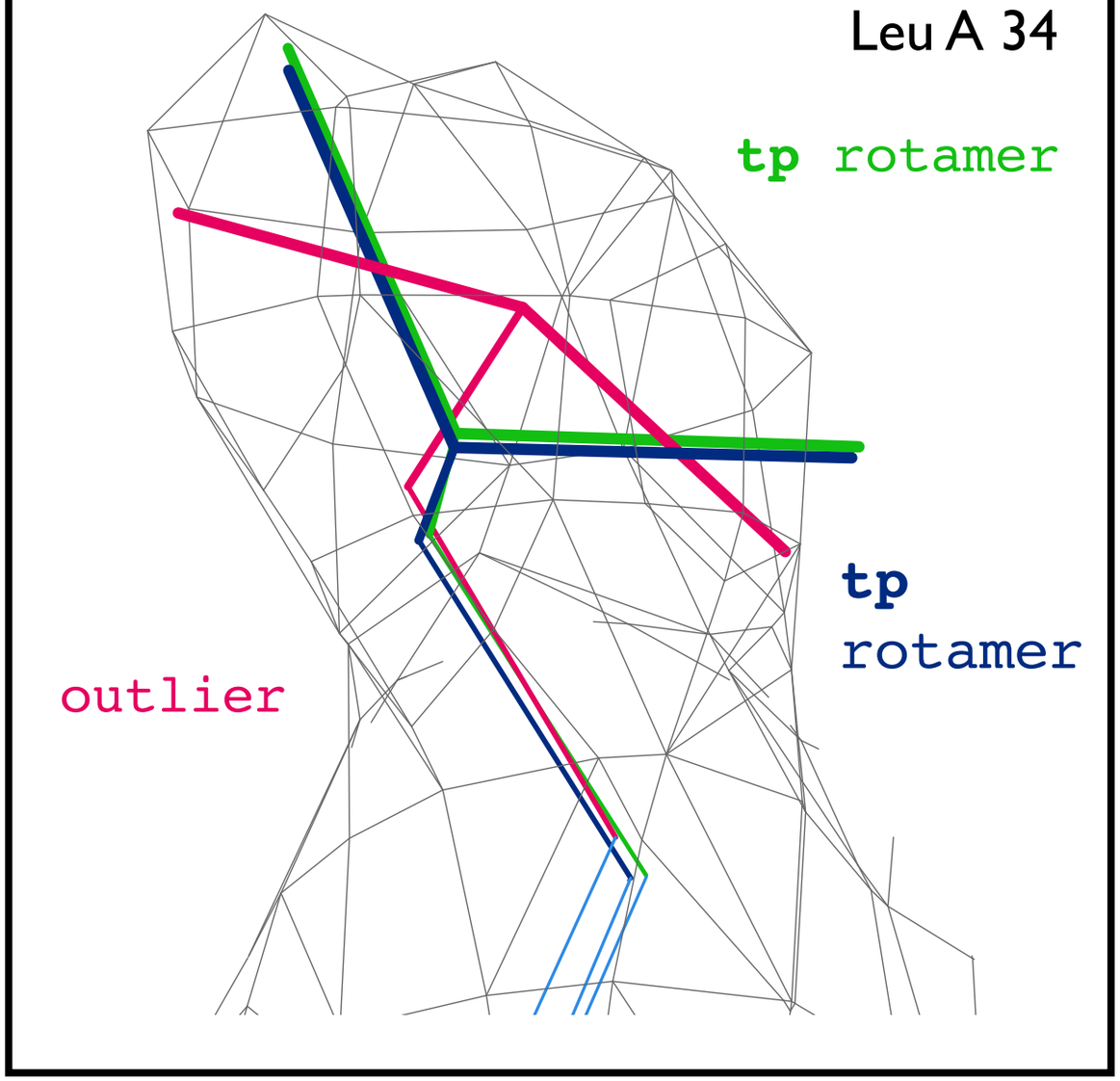
Why torsion angles?

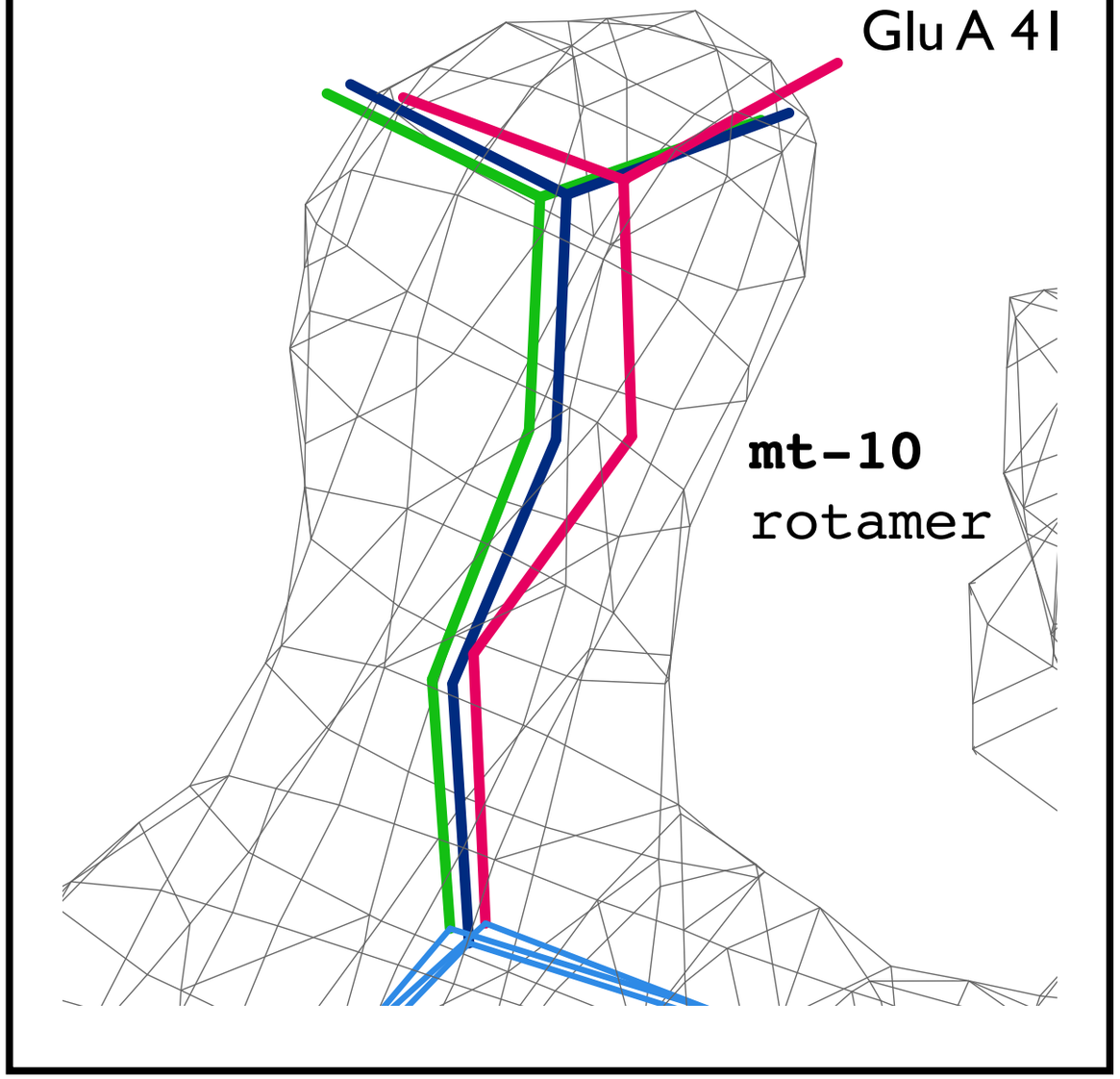


1GTX/1OHV reference example

outlier correction



restrained refinement



1GTX (3.0Å) 1OHV (2.3Å) 1GTX w/ 1OHV reference

5 macrocycles of *phenix.refine*
w/ reference restraints →

R_{free}: 0.2379 → 0.2186
ΔR: 0.833 → 0.60
MolProbity: 64th → 96th

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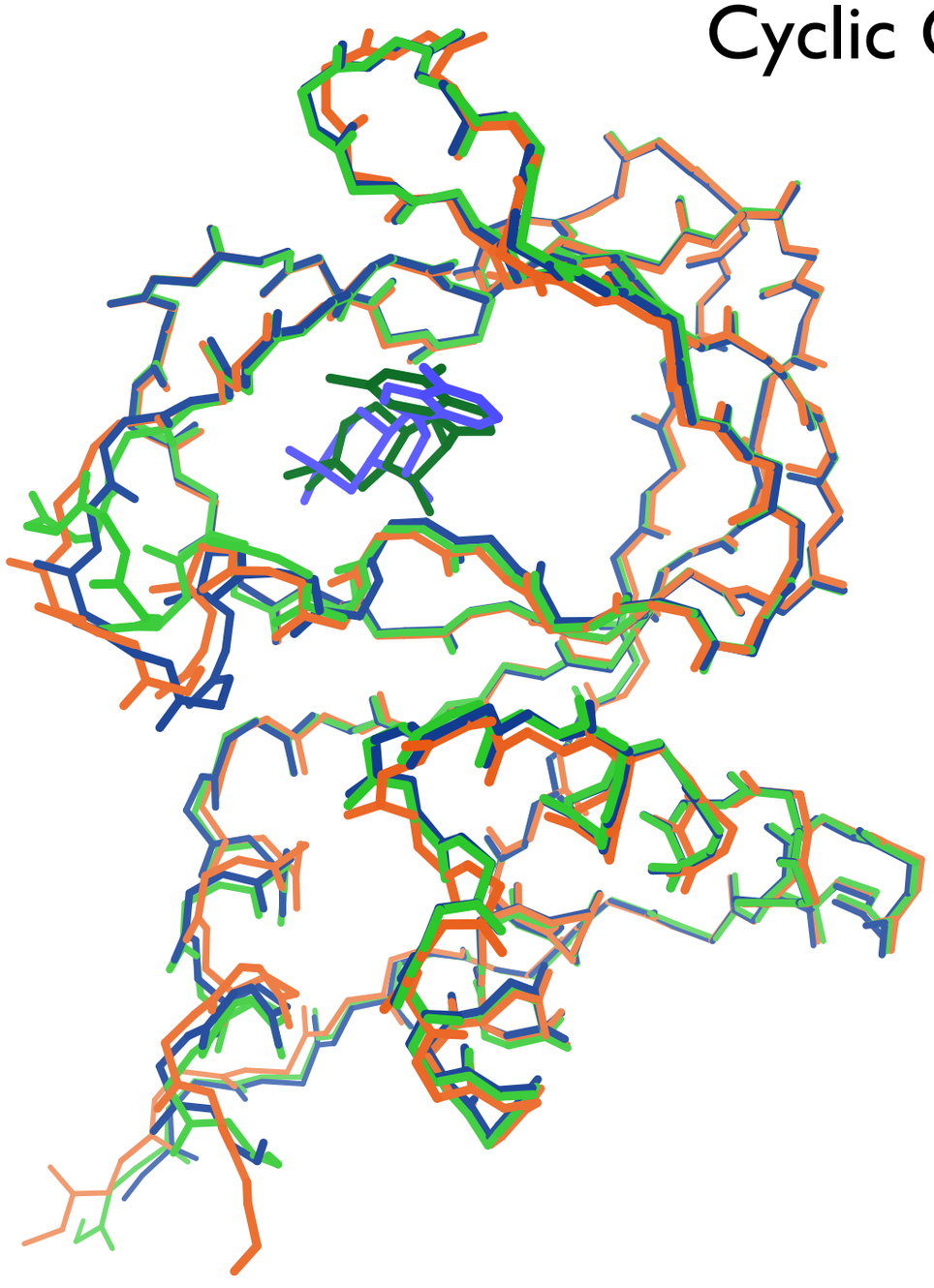
Practical Example

Cyclic GMP-dependent protein kinases (PKG's)



cAMP bound: 2.49Å

cGMP bound: 3.20Å

APO form: 2.69Å

JJ Kim et al. (2011) Crystal structures of PKG Iβ (92-227) with cGMP and cAMP reveal the molecular details of cyclic nucleotide binding. *PLoS ONE*.

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Cyclic GMP-dependent protein kinase

cAMP bound: 2.49Å

cGMP bound: 3.20Å

APO form: 2.69Å

	Validation Criteria	cAMP bound
All-Atom Contacts	Clashscore, all atoms:	16.53
	Clashscore percentile	81st
Protein Geometry	Poor rotamers:	2.61%
	Rama outliers:	0.00%
	Rama favored:	98.80%
	Cβ dev. > 0.25Å:	0
	MolProbity score:	2.04
	MP score percentile	95th
	Res w/ bad bonds:	0.00%
	Res w/ bad angles:	0.00%
Residual	R-work	0.1960
	R-free	0.2264

cGMP bound
56.57
15th
18.58%
2.02%
85.48%
23
3.84
12th
2.38%
5.95%
0.2102
0.2582

→

cGMP bound
24.56
87th
4.00%
0.40%
96.00%
0
2.61
96th
0.00%
1.18%
0.1980
0.2397

APO
28.52
46th
10.53%
3.19%
89.02%
3
3.29
12th
0.79%
0.98%
0.2205
0.2612

→

APO
19.5
15th
3.66%
0.60%
96.61%
0
2.43
89th
0.00%
0.20%
0.2166
0.2525

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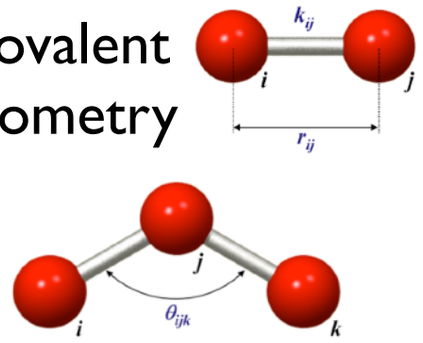
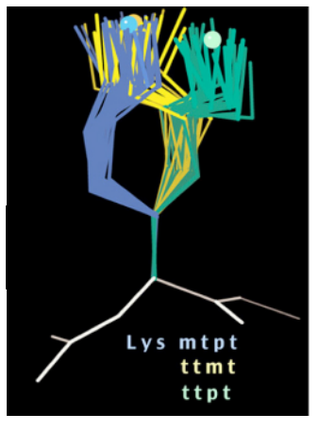
Sources of Prior Information

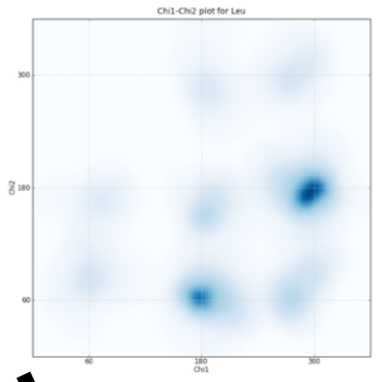
Covalent geometry

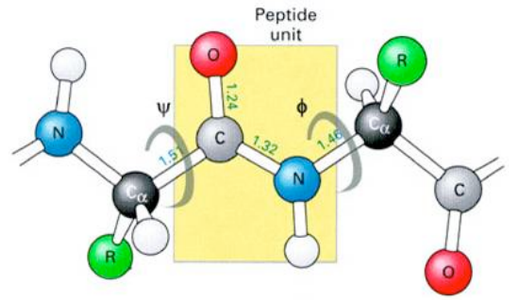


Images from PumMa web site (http://www.pumma.nl)

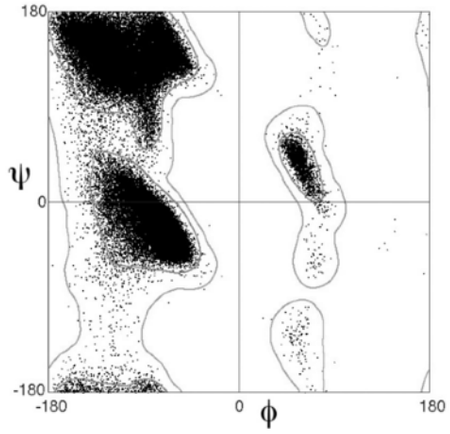
Sidechain distributions

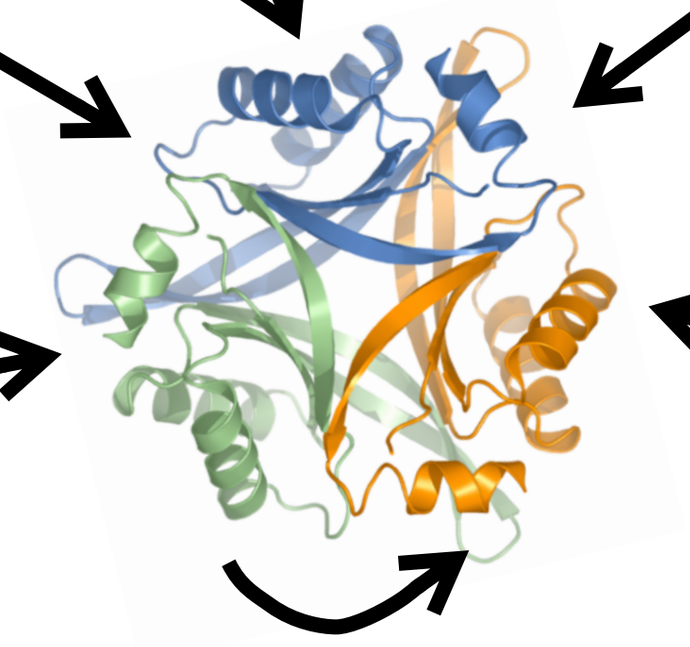


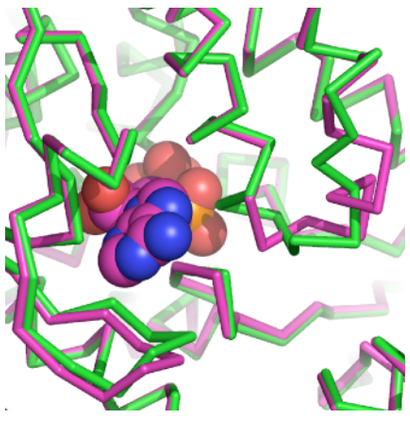




Mainchain distributions

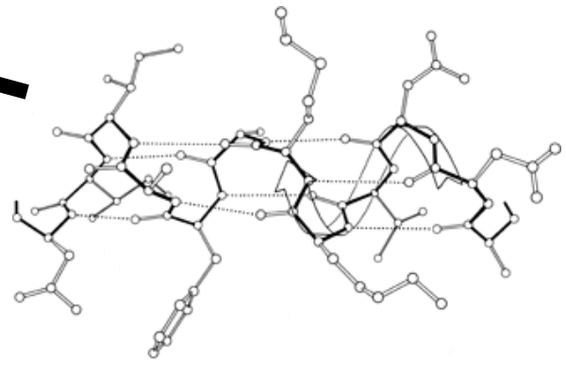






Related structures

Internal symmetry



Secondary structure

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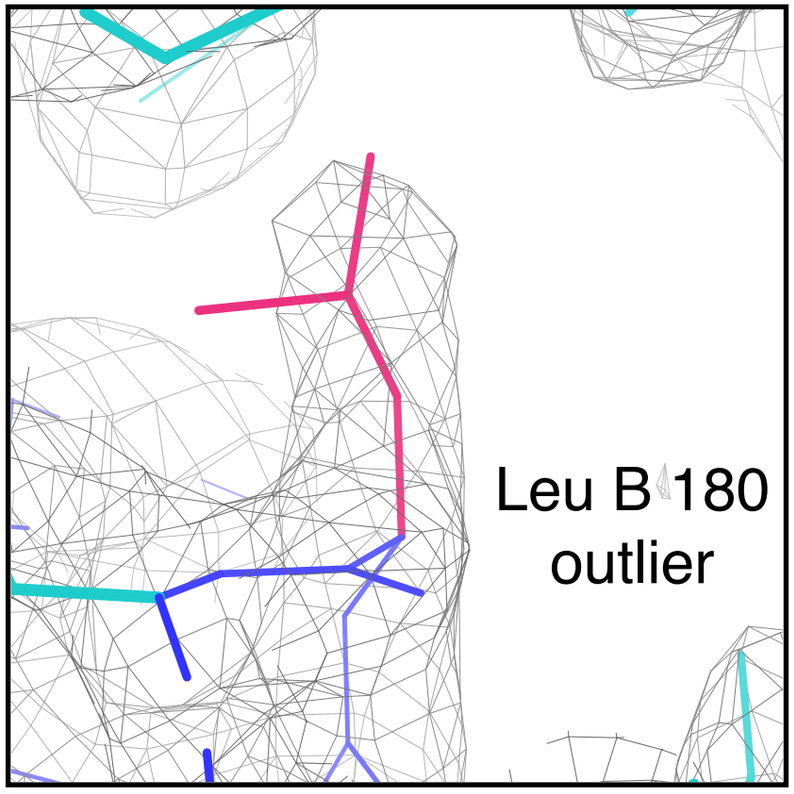
Torsion space NCS restraints

(Jeff Headd)

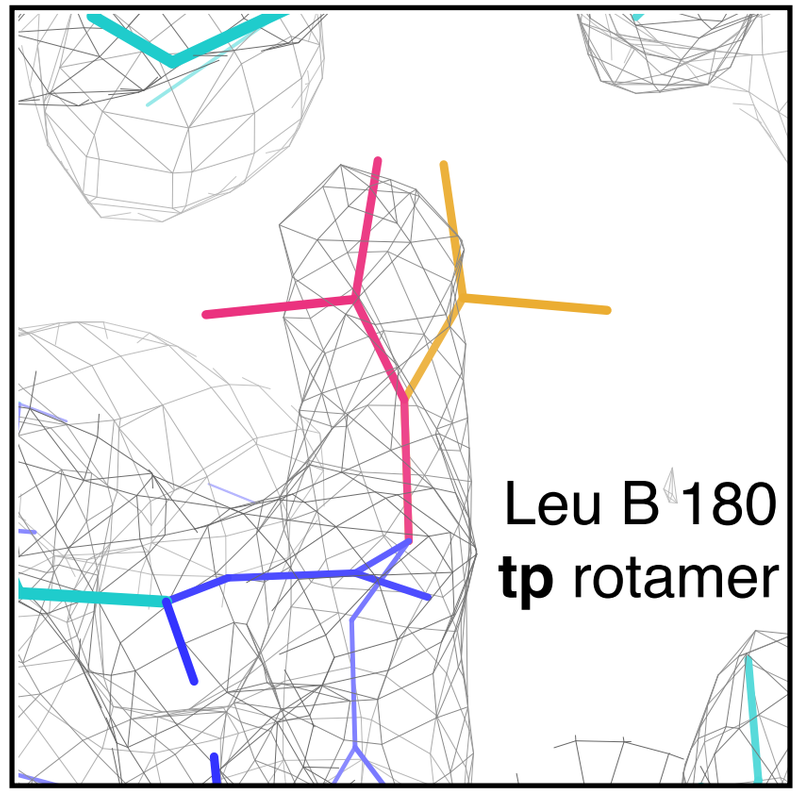
1b04: 2.8 Å DNA ligase

rotamer outlier correction

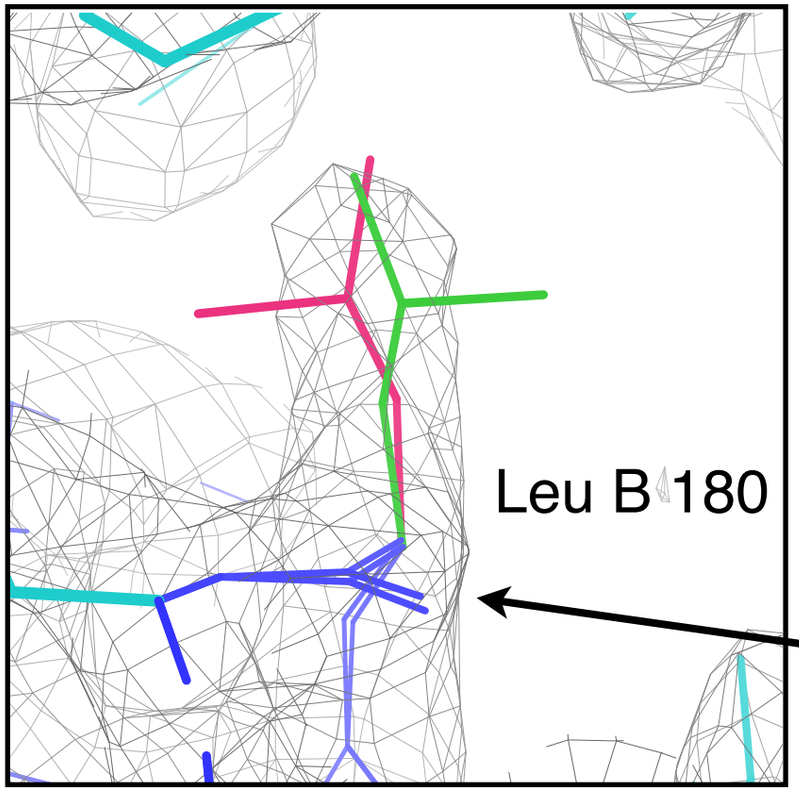
1. Identify rotamer outlier



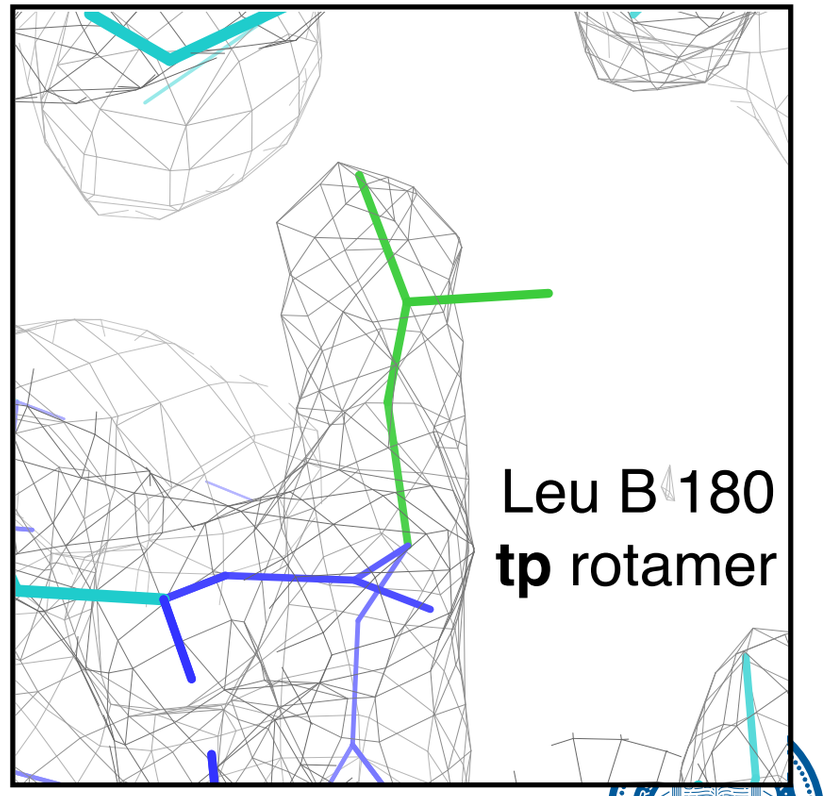
2. correct to corresponding rotamer in NCS-related chain by matching χ angles



3. ‘backrub’ search, then limited χ angle torsion search



4. verify rotamer is still correct match



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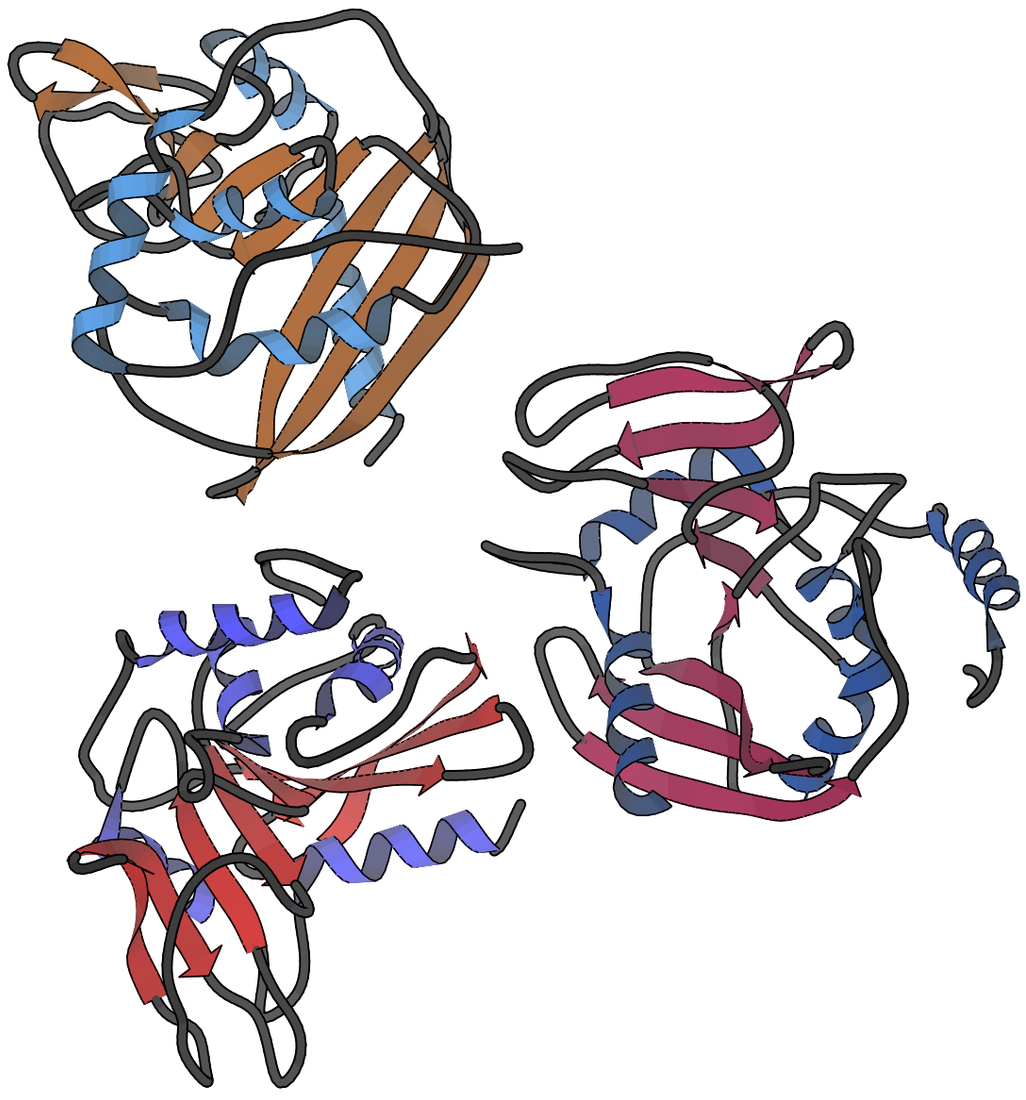
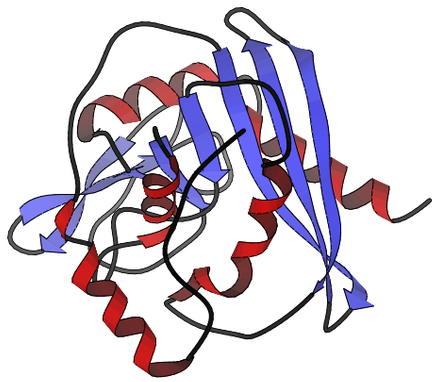
molecular replacement ⟶ refinement

3hd0: 2.70 Å
endonuclease



MR w/ Phaser



2w35
2.15 Å

R_{work} = 0.3844

AutoBuild

- Rebuild in place
- NCS on for rebuilding
- NCS off for refinement
- No water picking

R_{work} = 0.1895
R_{free} = 0.2745

R_{gap} = 0.085

phenix.refine

- 10 macrocycles
- optimize weights
- No NCS, Cartesian NCS, torsion NCS w/ and w/o rotamer correction

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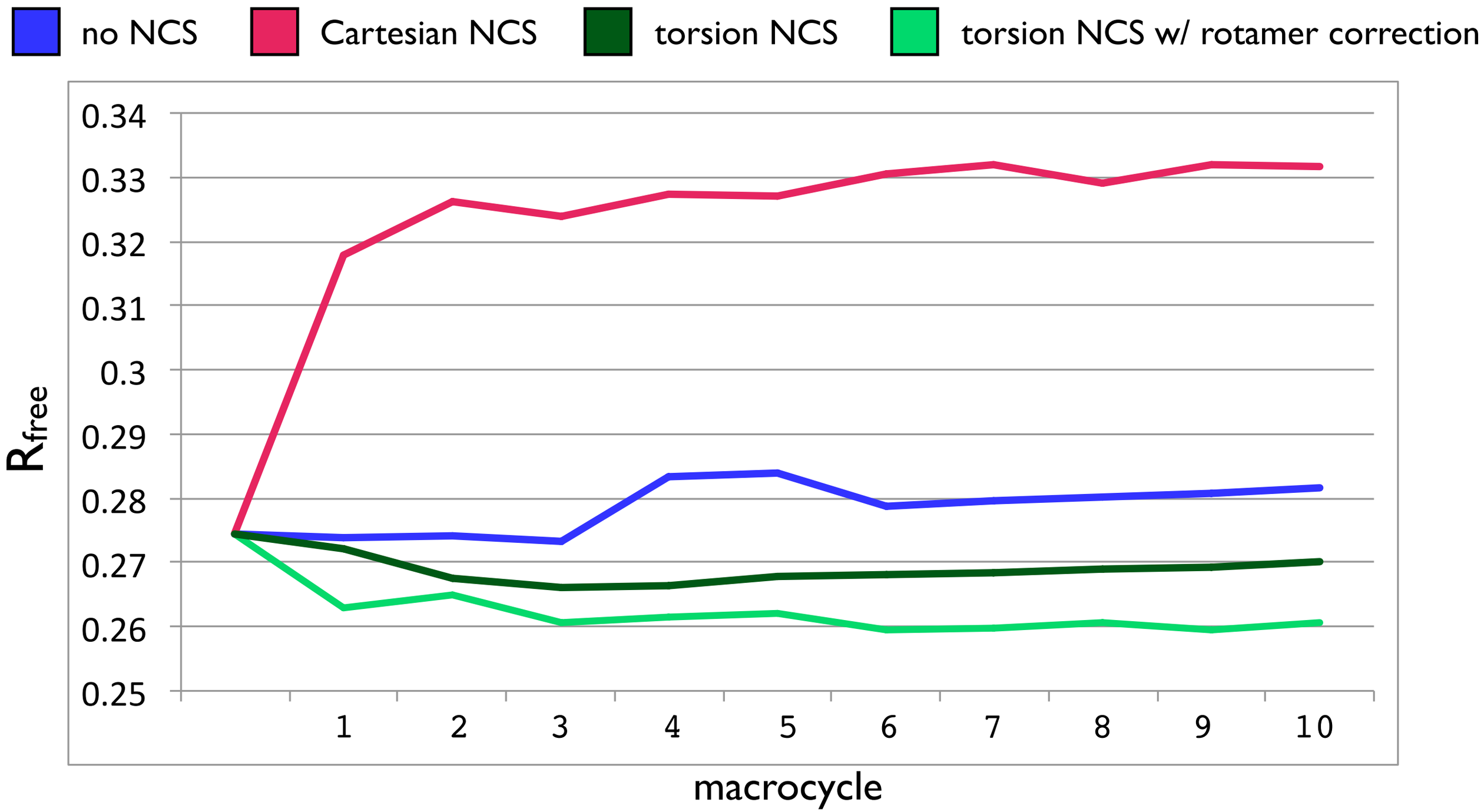
3hd0 refinement



torsion NCS w/ rotamer correction ⟶ $R_{work} = 0.2040$ $R_{free} = 0.2606$ $R_{gap} = 0.056$

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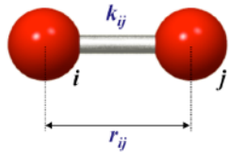
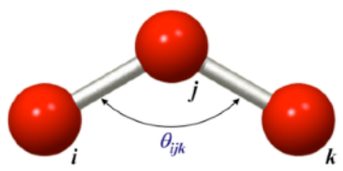
Sources of Prior Information

Covalent geometry





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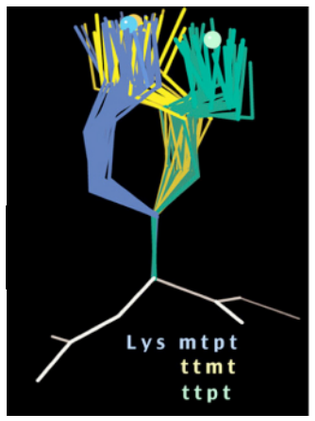
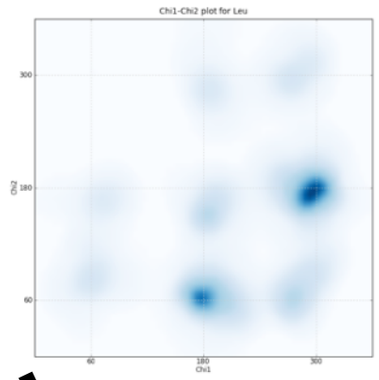
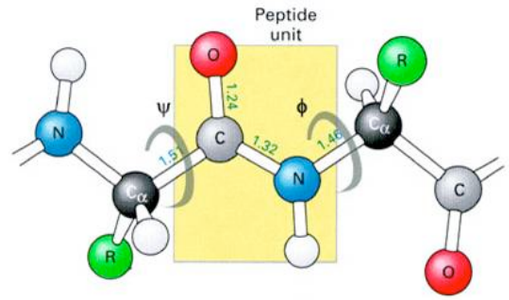
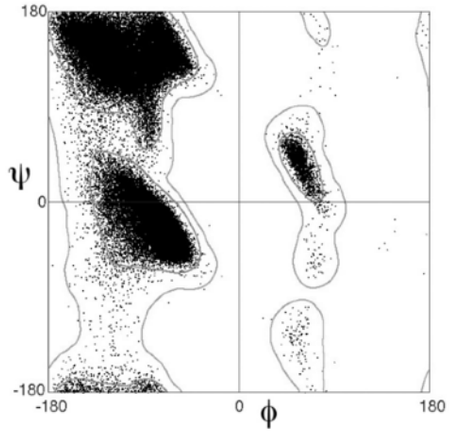
Sidechain distributions

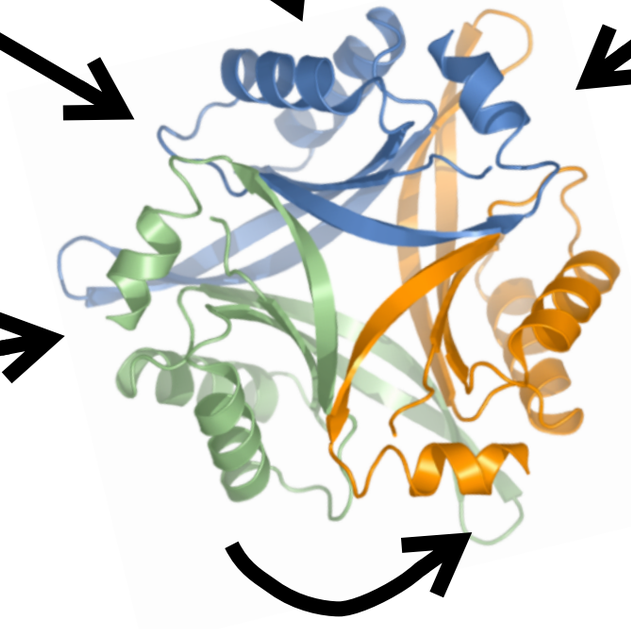


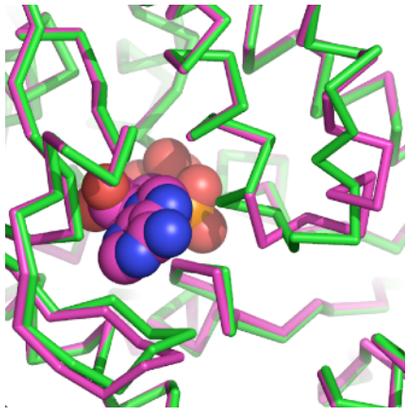




Mainchain distributions

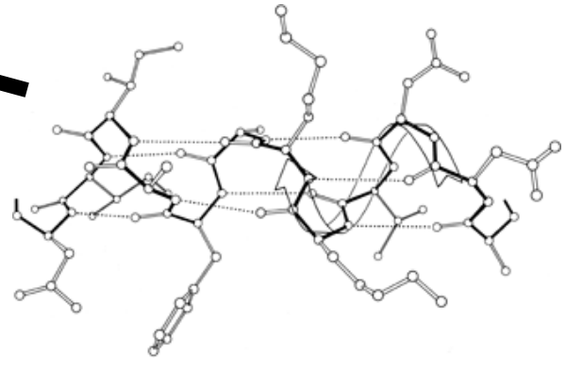






Related structures

Internal symmetry



Secondary structure

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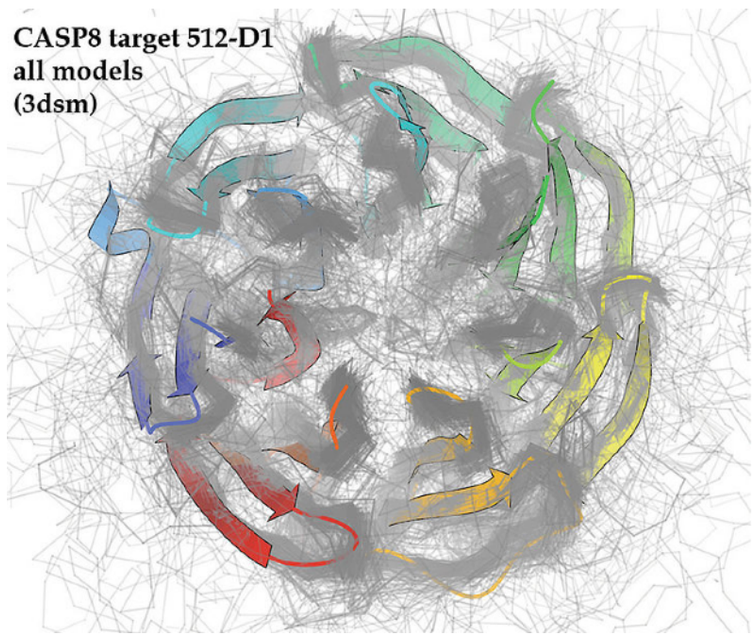
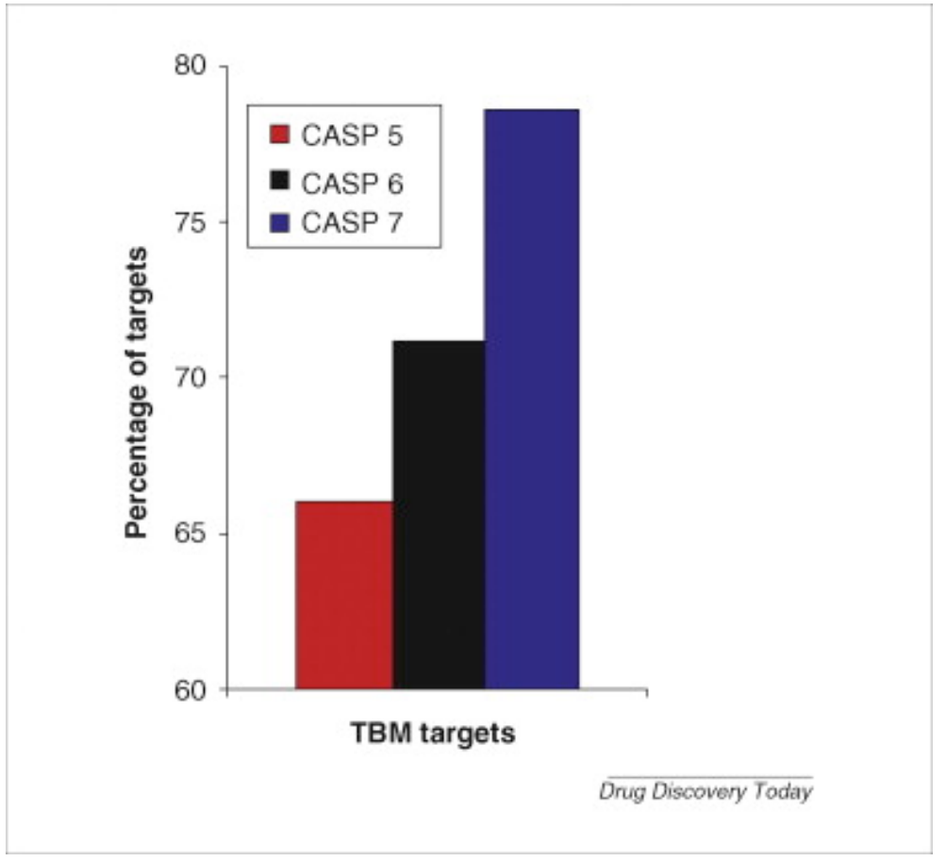
More Prior Information

- As the number of observations decreases we need to increase the amount of prior information we include (or the number of constraints we apply)
- At the extreme - what if we had no data?
- Other fields have been trying to address this problem:
  - Structure prediction
  - Homology modelling
  - Protein folding



http://www.predictioncenter.org



From: Kryshtafovych & Fidelis, Drug Discovery Today, 2009, 14:386–393

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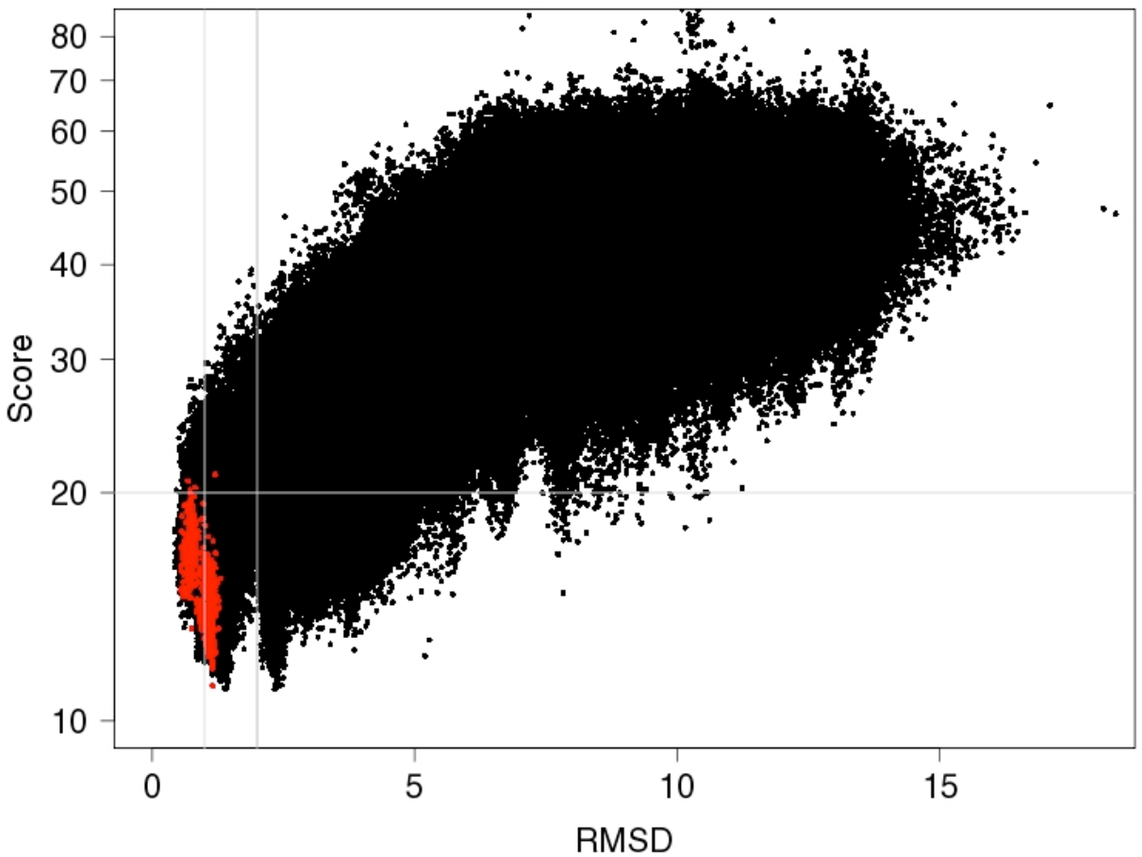
Physically Realistic Potentials (Rosetta)

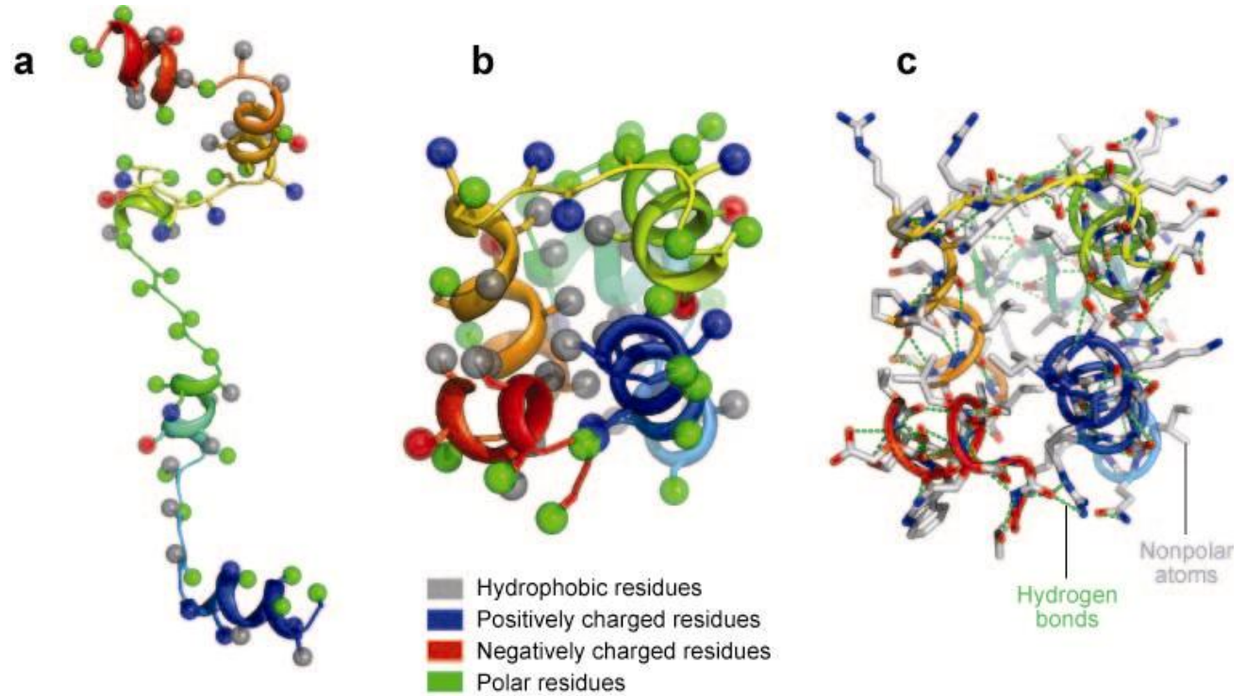
(Nat Echols & Frank DiMaio)

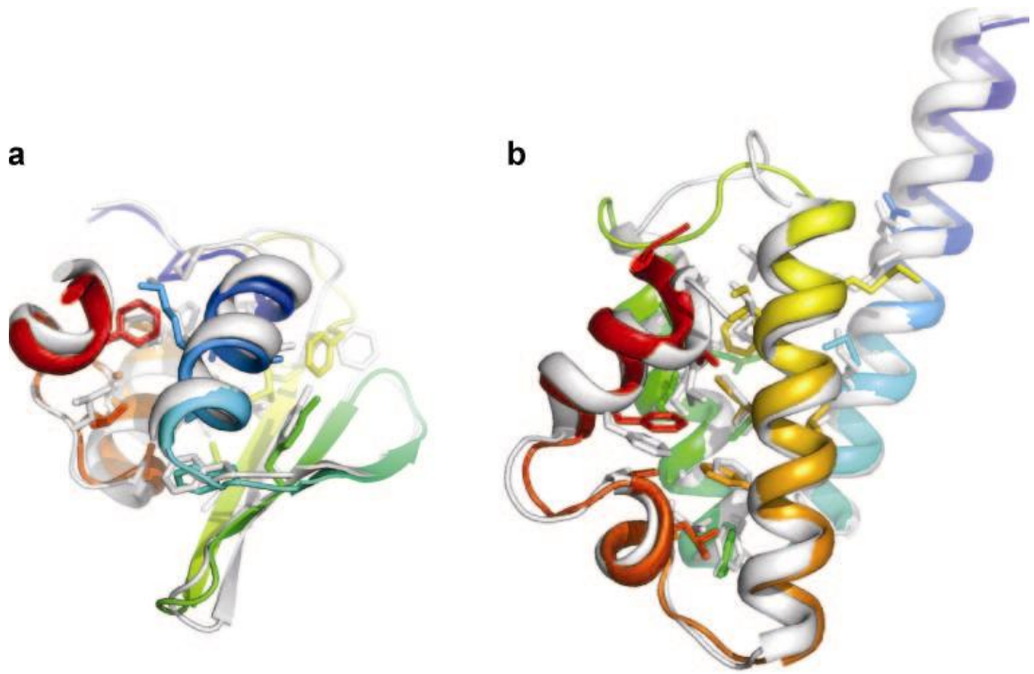
Rosetta

- *ab initio* model generation and model optimization
- Requires extensive computational sampling



Black - Rosetta *ab initio* models, Red - Crystal structure after Relax protocol





Das R, Baker D. 2008.
Annu. Rev. Biochem. 77:363–82.

Why Rosetta

- Designed to recognize near-native structures among many possible models; combines empirical and physical potentials
  - All-atom force field, incorporates solvation effects, attractive forces, hydrogen bonds, knowledge-based dihedral restraints
- Can yield chemically realistic *ab initio* models without experimental data to guide assembly
  - Occasionally good enough for molecular replacement
- Shown to be useful for NMR structure determination with sparse data (CS-Rosetta), MR solution improvement (MR-Rosetta), RNA structure refinement (ERRASER)

Kuhlman et al. (2003) *Science* **302**:1364-8
Rohl et al. (2004) *Methods Enzymol.* **383**:66-93
Keedy et al. (2009) *Proteins* **77**:29-49

https://www.rosettacommons.org

Complementary Algorithms

Phenix

- Reciprocal space X-ray target functions (ML, MLHL, LS-twin)
- Bulk solvent correction
- B-factor refinement (including TLS)
- Map calculation
- Density modification (using RESOLVE)

Rosetta

- Physically realistic potentials
- Repacking to remove steric clashes and building rotameric sidechains
- Torsion-angle minimization
- Real-space target (refinement against electron density)
- Fragment-based rebuilding (optional, not currently used)

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Python/C++ architecture facilitates combination

Low Resolution Protocol

- Sidechain repacking (using density)
- Coordinate refinement (reciprocal space torsion angle minimization and reduced nonbonded penalty)
- B-factor refinement

3 Cycles

- Sidechain repacking (using density)
- Coordinate refinement (real space and reciprocal space torsion angle minimization)
- B-factor refinement

5 Cycles

- Sidechain repacking (using density)
- Coordinate refinement (reciprocal space minimization with restrained bonds and angles)
- B-factor refinement

2 Cycles

Protocol run 5 times in parallel and the best model selected based on R-free

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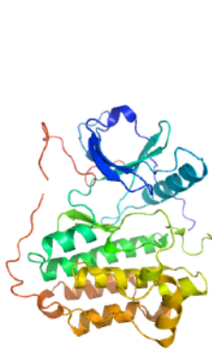
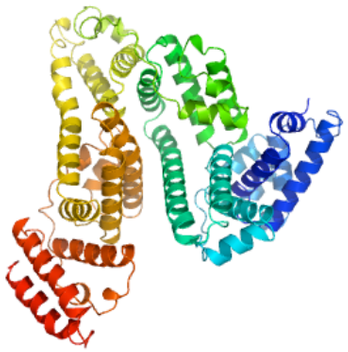
Test Cases

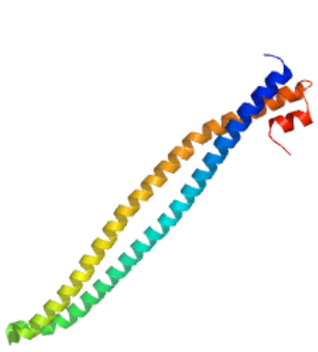
Solved by molecular replacement with same protein from another deposition at higher resolution



2j5f (3.0Å)



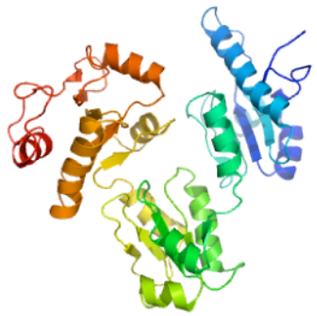
1bke (3.15Å)



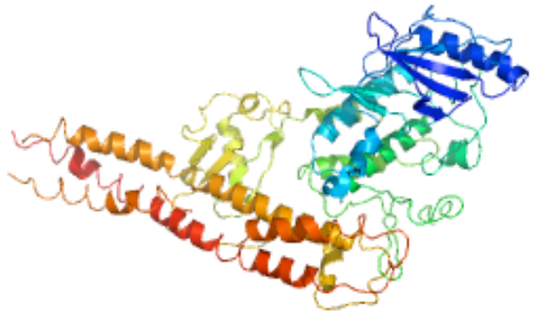
3mtt (3.3Å)

1kct (3.46Å)

3snh (3.7Å)

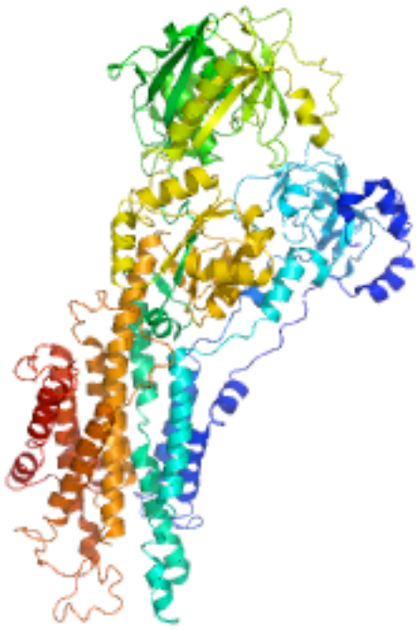


2vaf (3.8Å)

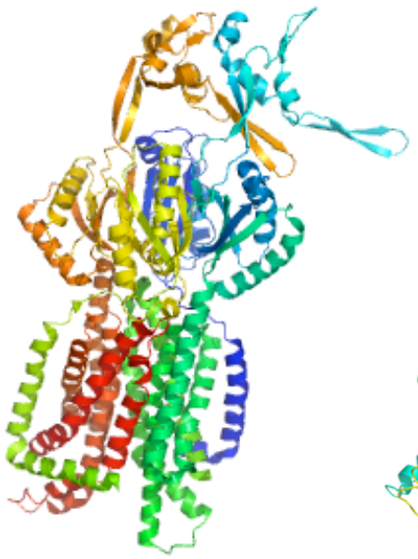


3rzf (4.0Å)

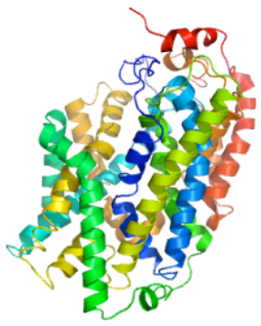
Membrane Proteins



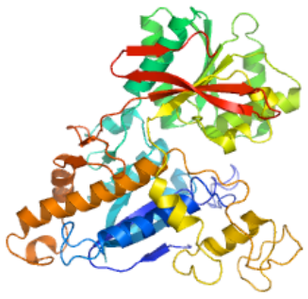
3fps (3.2Å)



3k07 (3.52Å)

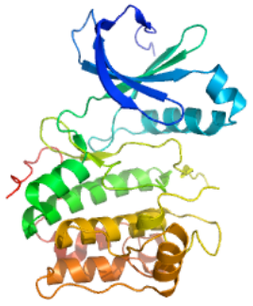


2x79 (3.8Å)

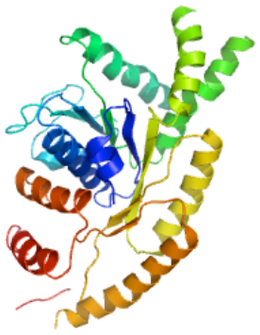


1isr (4.0Å)

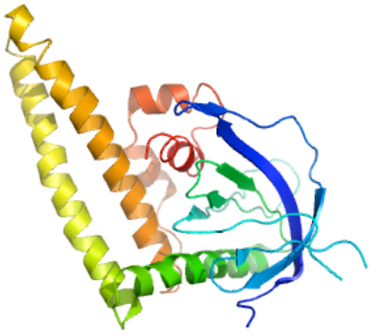
Solved using homologous proteins



3pwy (3.5Å)



3idq (3.7Å)



3a8n (4.5Å)

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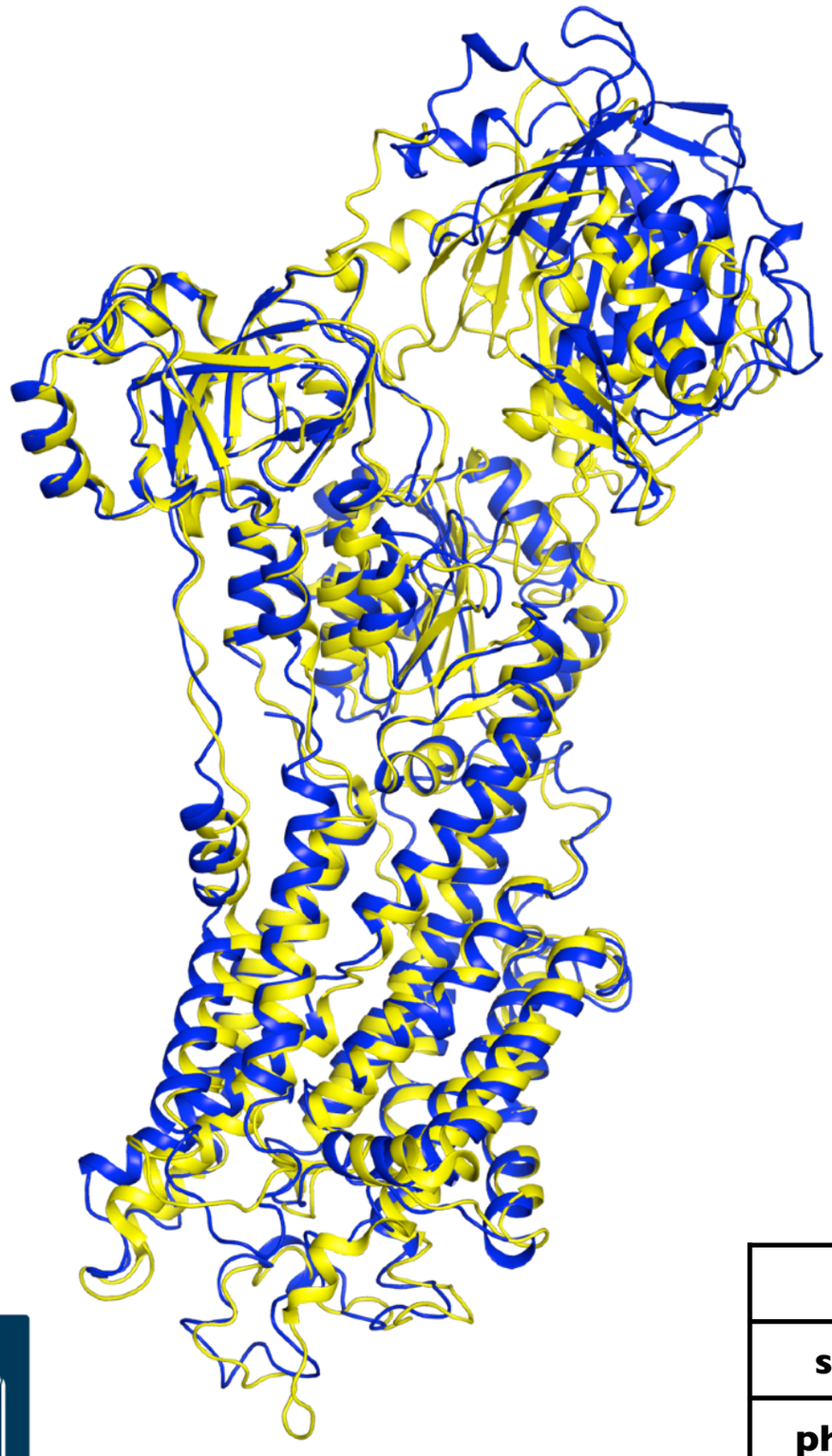
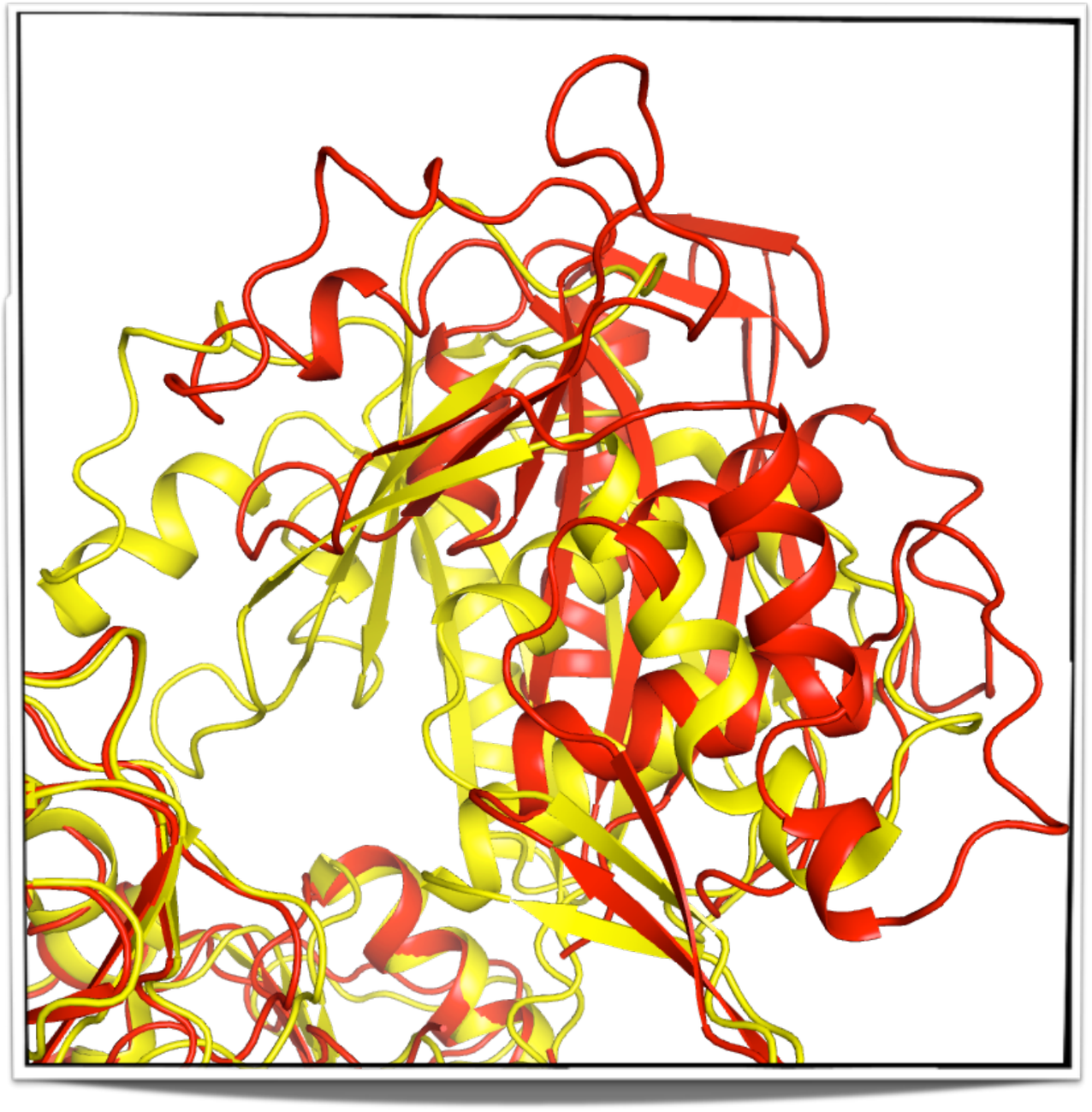
Calcium ATPase - phenix.refine





	R	R-free	mp score	RMSD
start	0.47	0.51	3.21	6.1
phenix	0.43	0.48	2.66	6.2

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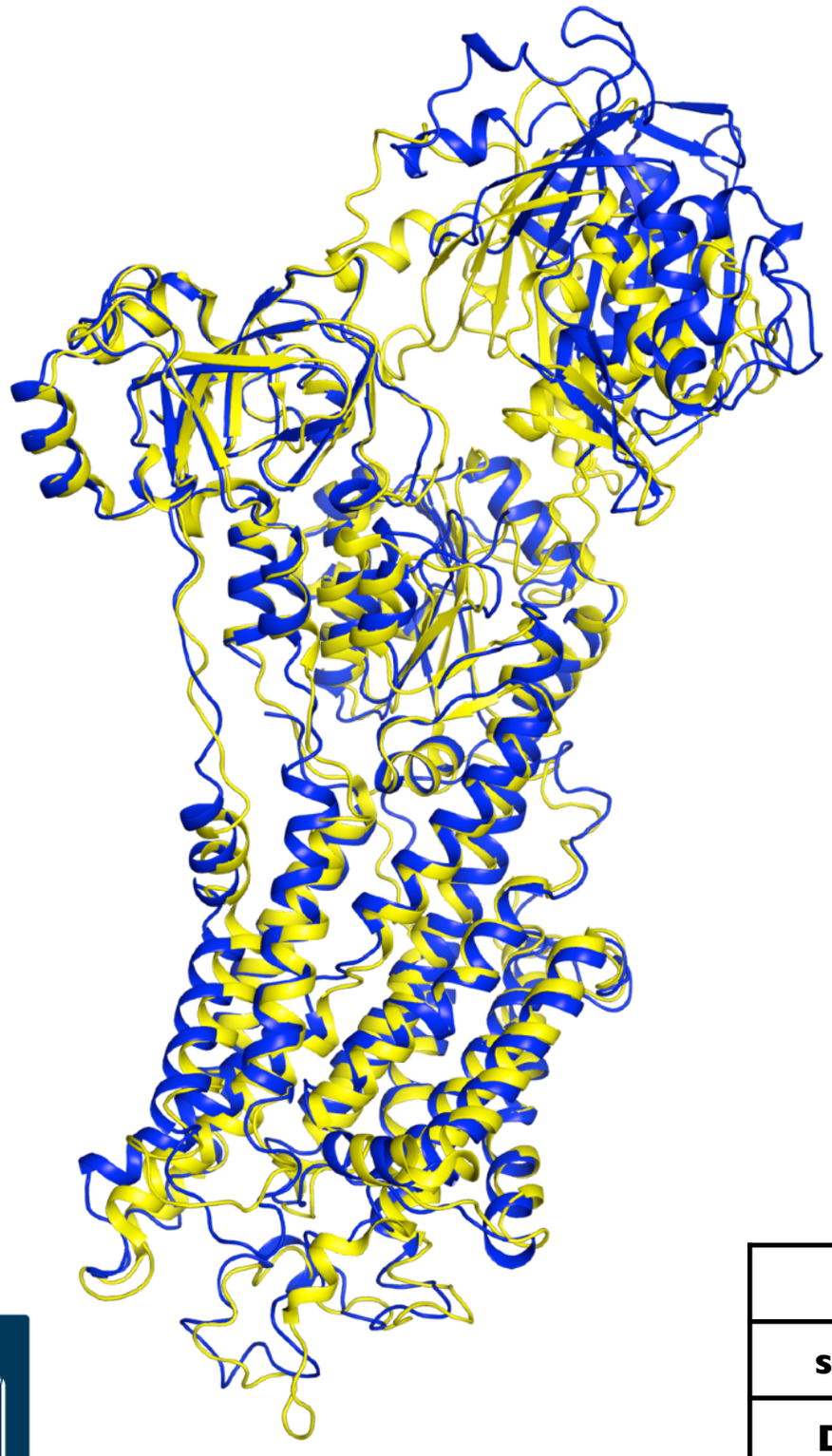
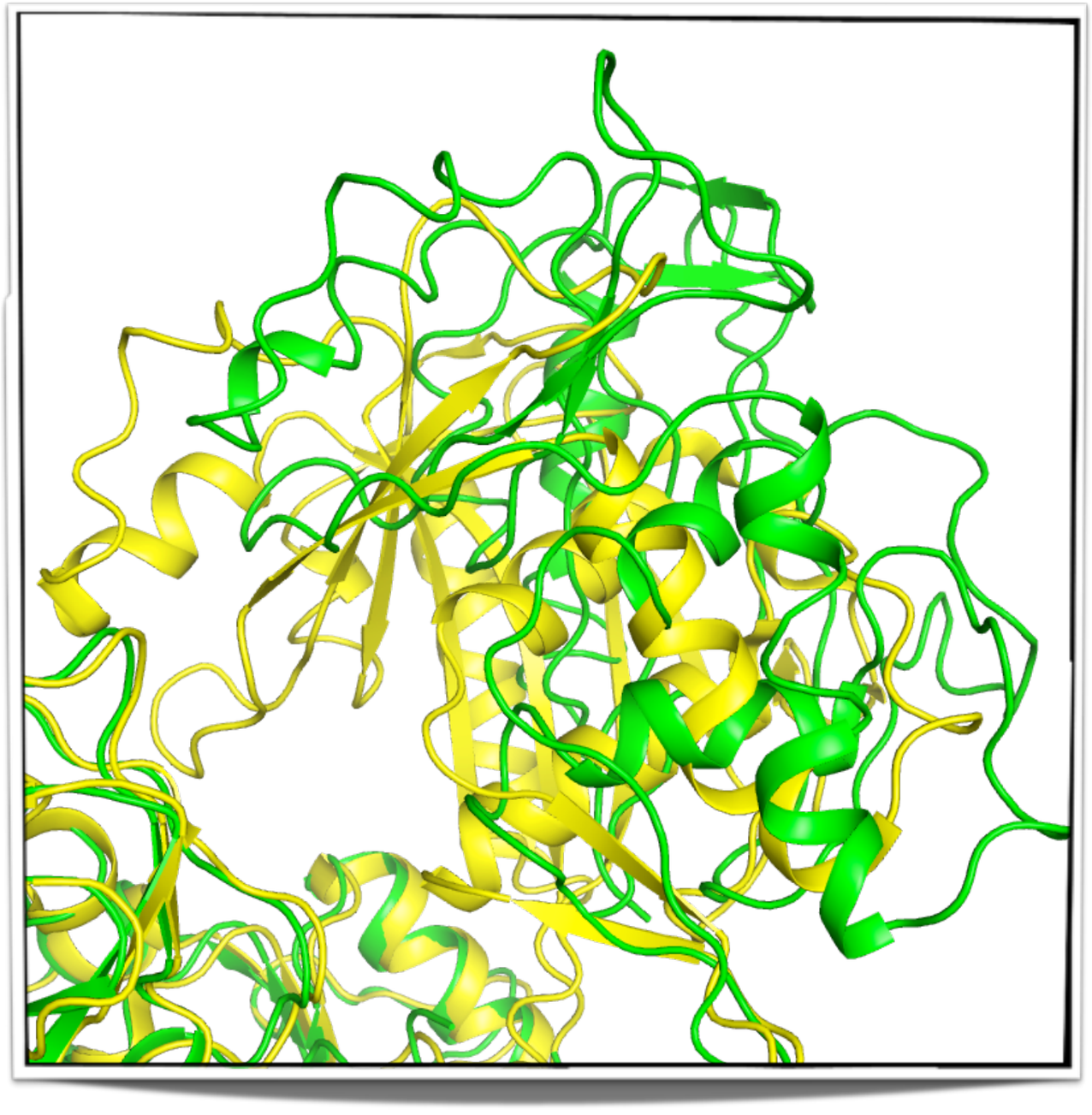
Calcium ATPase - DEN





	R	R-free	mp score	RMSD
start	0.47	0.51	3.21	6.1
DEN	0.38	0.44	3.79	6.1

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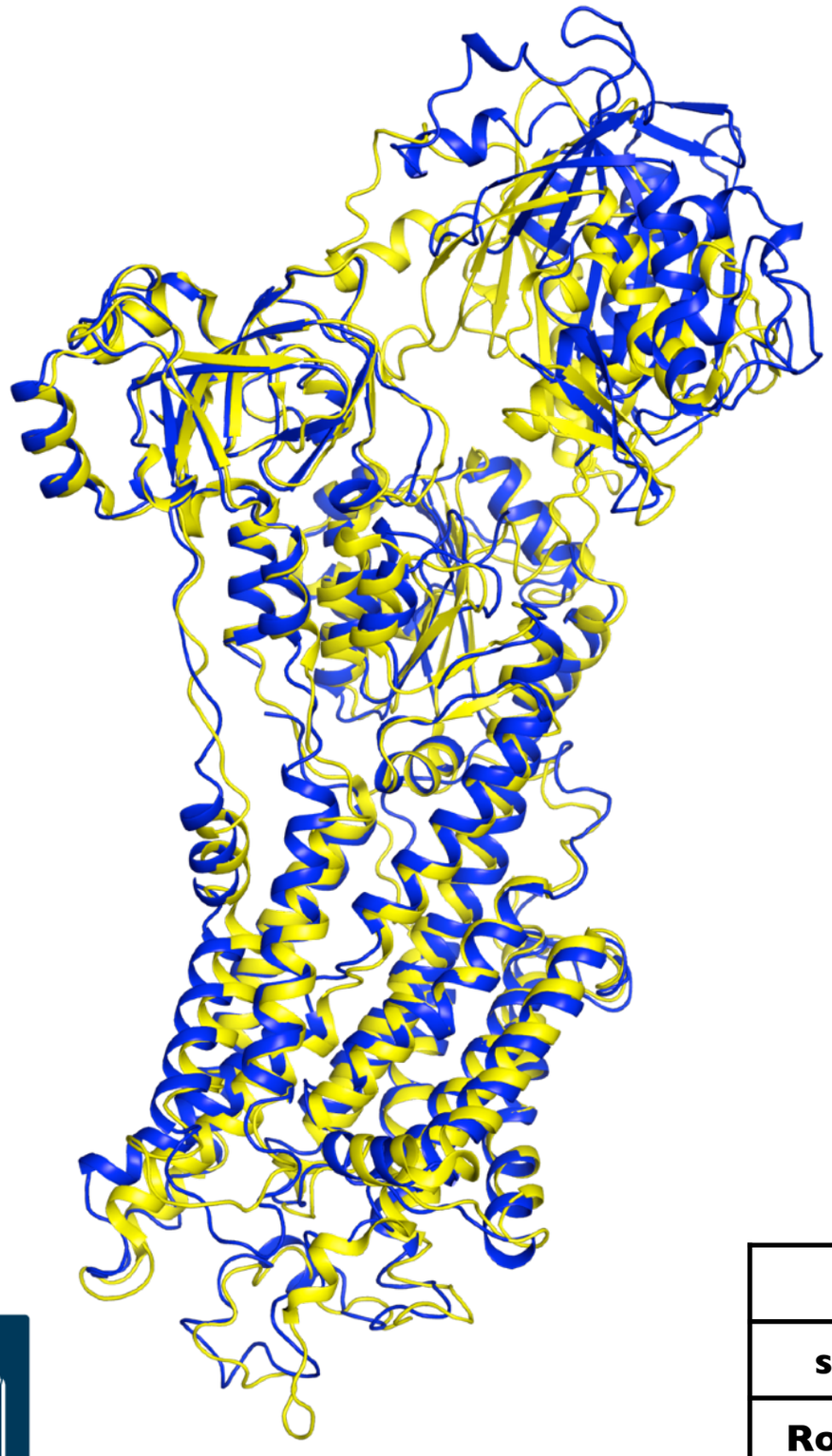
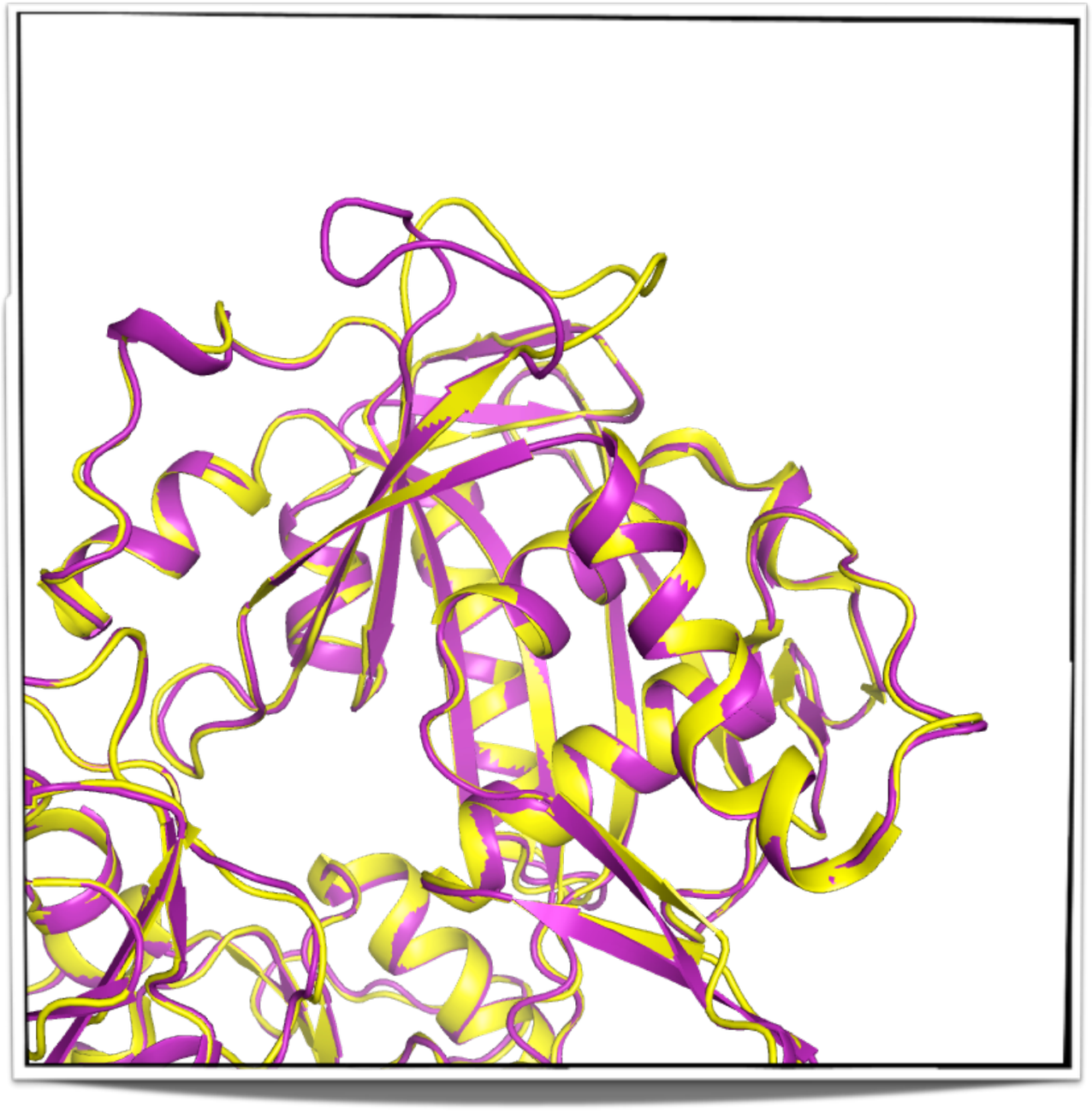
Calcium ATPase - Phenix-Rosetta





	R	R-free	mp score	RMSD
start	0.47	0.51	3.21	6.1
Rosetta	0.24	0.28	1.55	1.7

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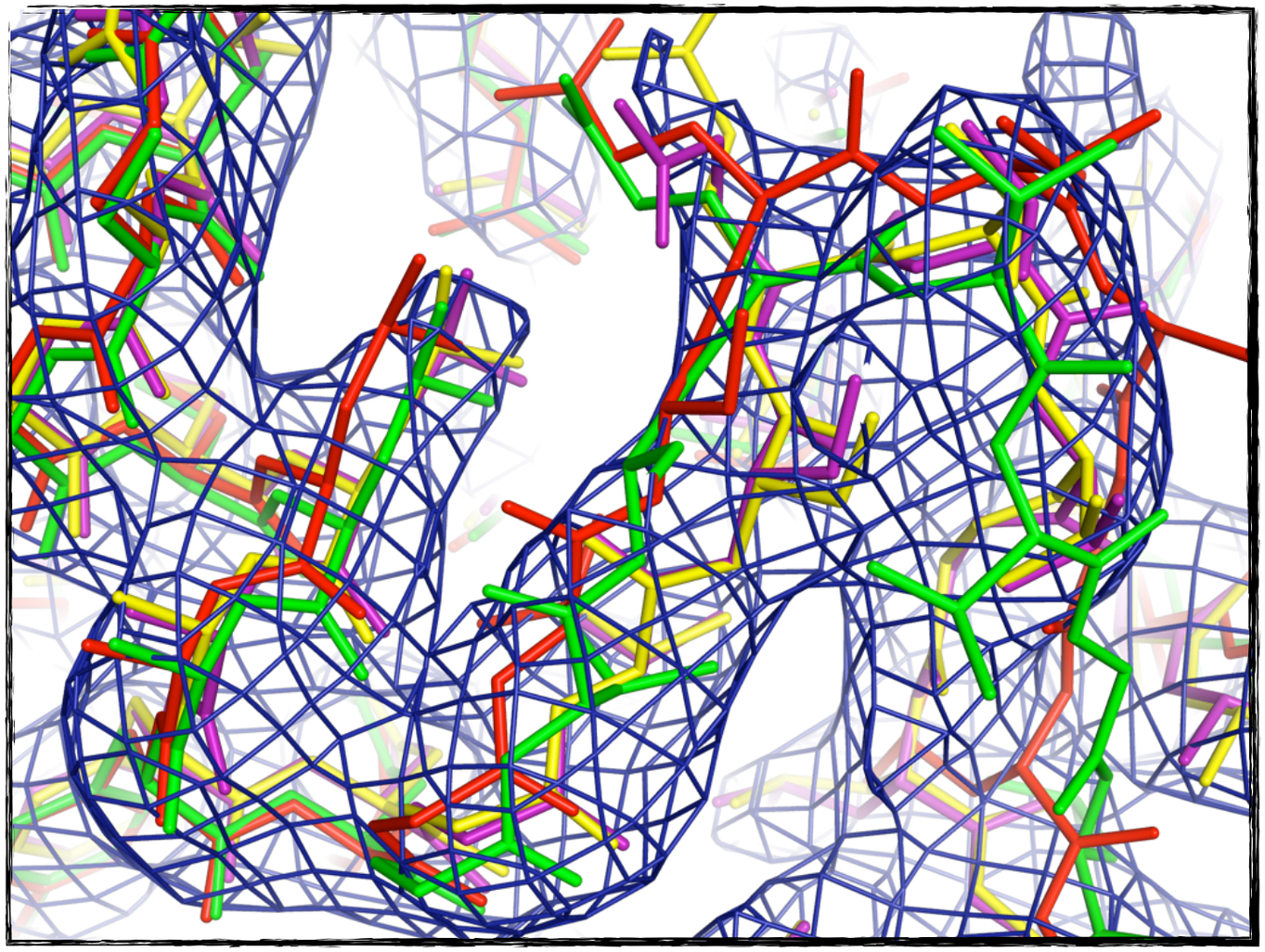
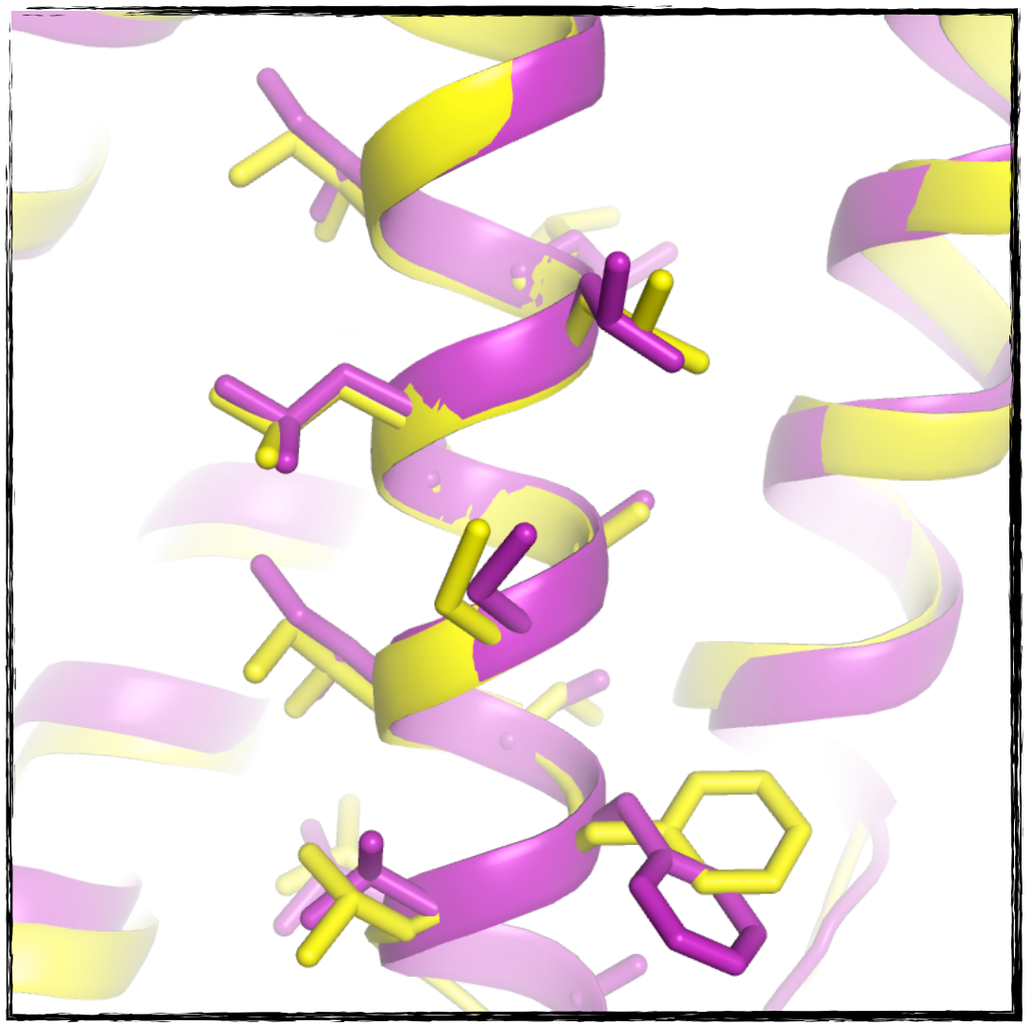
Calcium ATPase - Detail





- Phenix-Rosetta model is very close to the deposited structure (even at the level of side chains) with better fit to density

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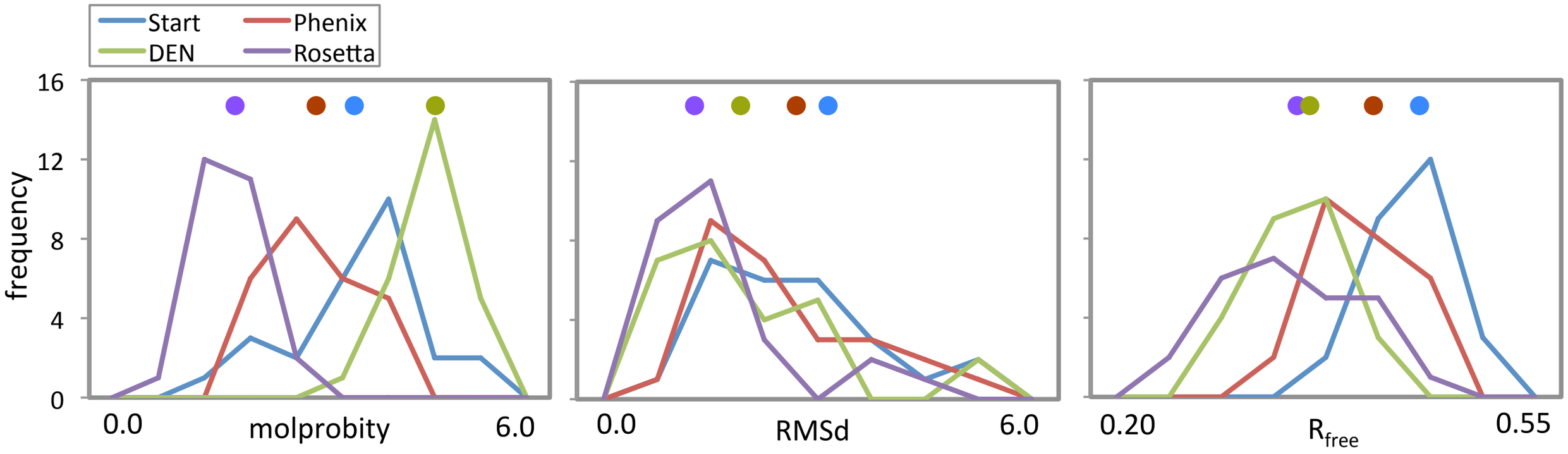
Improved Models



- Phenix-Rosetta typically has improved fit to the crystallographic data and models are closer to the known structure
- Phenix-Rosetta always has improved model quality, as judged by Molprobity
- Generally similar to DEN results but with much improved geometry, and generally faster

DiMiao et al., 2013, *Nature Methods* ***10***:*1102-1104*

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- **Lawrence Berkeley Laboratory**
  - Pavel Afonine, Youval Dar, Nat Echols, Jeff Headd, Richard Gildea, Ralf Grosse-Kunstleve, Dorothee Liebschner, Nigel Moriarty, Nader Morshed, Billy Poon, Ian Rees, Nicholas Sauter, Oleg Sobolev, Peter Zwart
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- **Duke University**
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- **Oak Ridge National Laboratory**
  - Marat Mustyakimov, Paul Langan
- **Others**
  - Alexandre Urzhumtsev & Vladimir Lunin
  - Garib Murshudov & Alexi Vagin
  - Kevin Cowtan, Paul Emsley, Bernhard Lohkamp
  - David Abrahams
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  - Lawrence Berkeley Laboratory
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